INSTRUCTIONS

T-Gel[™] Purification Kit



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44916

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Number	Description
44916	T-Gel [™] Purification Kit
	Kit Contents:
	T-Gel[™] Adsorbent Columns, 4 x 3 ml, contains T-Gel [™] Storage Buffer
	Support: 6% beaded agarose
	Bead Diameter: 45-165 µm
	Binding Capacity: ~20 mg of human IgG/ml of settled gel
	T-Gel[™] Binding Buffer, 1 L, contains 0.5 M potassium sulfate, 50 mM sodium phosphate, 0.05% sodium azide; pH 8.0
	T-Gel [™] Elution Buffer, 1 L, contains 50 mM sodium phosphate, 0.05% sodium azide; pH 8.0
	T-Gel TM Column Storage Buffer (2X), 100 ml, contains 1 M Tris and 0.05% sodium azide; pH 7.4
	Guanidine Hydrochloride, 230 g, sufficient reagent to prepare 300 ml of an 8 M solution
	Column Extenders, 4 each
	Storage: Upon receipt store product at 4°C. Do not allow gel to freeze. Product is shipped at ambient temperature.

Warranty: Pierce products are warranted to meet stated product specifications and to conform to label descriptions when used and stored properly. Unless otherwise stated, this warranty is limited to one year from date of sale for products used, handled and stored according to Pierce instructions. Pierce's sole liability for the product is limited to replacement of the product or refund of the purchase price. Pierce products are supplied for laboratory or manufacturing applications only. They are not intended for medicinal, diagnostic or therapeutic use. Pierce products may not be resold, modified for resale or used to manufacture commercial products without prior written approval from Pierce Biotechnology.

Introduction

T-GelTM Adsorbent allows for simple, rapid, one-step immunoglobulin purification from a wide variety of serum, ascites or tissue culture supernatant samples. Immunoglobulin purification using T-GelTM Adsorbent is based on the ability of some proteins to bind to a ligand that contains a sulfone group in proximity to a thioether group (Figure 1). This binding event is termed thiophilic adsorption and is a highly selective type of lyotropic salt-promoted interaction.

Thiophilic adsorption has some elements of both hydrophobic and hydrophilic adsorption. Increased non-chaotropic salts promote both thiophilic and hydrophobic interactions. However, hydrophobic interaction chromatography is strongly promoted by high concentrations of sodium chloride, whereas thiophilic adsorption is not. Salts that interact with water molecules, such as potassium sulfate and ammonium sulfate, promote protein binding to thiophilic supports.

T-GelTM Adsorbent has a high binding capacity and a broad specificity toward immunoglobulins from various species regardless of the immunoglobulin type or subclass. This method provides a low cost, efficient alternative to ammonium sulfate precipitation as the first step of a multi-step immunoglobulin purification scheme for crude samples. The T-GelTM Adsorbent exhibits high protein recovery with excellent preservation of antibody activity. The gentle elution conditions yield concentrated, essentially salt-free, highly purified immunoglobulins at near-neutral pH. Thus, this simple one-step method eliminates the need for additional treatment of the sample for storage or for subsequent conjugation reactions.

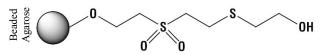


Figure 1. Molecular structure of the T-Gel[™] Adsorbent immobilized ligand. Telephone: 800-8-PIERCE (800-874-3723) or 815-968-0747 • Fax: 815-968-7316 or 800-842-5007 www.piercenet.com • Customer Service: cs@piercenet.com



Important Product Information

- Temperature, pH, ionic strength and specific salts affect binding and elution efficiency and sample purity. High concentrations of non-chaotropic salts improve coupling efficiency, but chaotropic salts that do not form structures with water decrease coupling efficiency.
- Coupling at pH<8 will generally increase protein binding; however, greater amounts of proteins other than immunoglobulins will also bind to the support.
- When 1 ml of sera is applied to a 3 ml column of T-Gel[™] Adsorbent, essentially all immunoglobulins present will bind. However, at larger sample volumes, one or more of the non-bound (NB) fractions will contain immunoglobulins. These NB fractions may be pooled and treated as a sample for a subsequent purification to recover all immunoglobulins from the original sample.

Additional Materials Required

• Crystalline Potassium sulfate, ACS Reagent Grade

Material Preparation

Sample Preparation	While mixing, add 87 mg of potassium sulfate per milliliter of sample for a final concentration of 0.5 M potassium sulfate. Gently mix sample to avoid denaturation of the immunoglobulins. When the potassium salt is fully dissolved, centrifuge sample at 10,000 x g for 20 minutes. Carefully remove the clear supernatant and filter it using a 0.5 μ m filter.
Regeneration Solution	Add 124 ml ultrapure water to 230 g of crystals to prepare 300 ml of the 8 M reagent. Stir at room temperature until completely dissolved. Solubilization of guanidine•HCl is endothermic and may require mild warming in a 37°C water bath to completely dissolve. Allow the reagent to equilibrate to room temperature before using. Store solution 4°C for up to one year.

Procedure for Immunoglobulin Purification using T-Gel[™] Adsorbent

- 1. Equilibrate T-GelTM Adsorbent, buffers and sample(s) to room temperature.
- 2. To prevent air bubbles from forming in the gel bed and below the porous discs, open a T-Gel[™] Adsorbent column by first removing the top cap and then the bottom cap. Allow the storage buffer to drain.
- 3. Equilibrate column with 12 ml of Binding Buffer.
- 4. Apply the sample to the column and allow the sample to completely enter the gel. The column flow will cease when the liquid level reaches the top disc. If desired, collect the column effluent as 3 ml non-bound (NB) fractions.
- 5. Wash the column with up to 13 column volumes of Binding Buffer. Monitor absorbance of the fractions at 280 nm to determine when all NB material is washed from the column.
- 6. Elute immunoglobulins with 12 column volumes of Elution Buffer and collect the effluent as 3 ml fractions. Measure the absorbance of each fraction at 280 nm vs. water.
- 7. Regenerate the T-Gel[™] Adsorbent by adding five column volumes of Regeneration Solution to the column and allowing the column to drain. Rinse column with 10 column volumes of degassed ultrapure water followed by two column volumes of storage buffer. Allow the column to drain.

Note: The Regeneration Solution completely removes all residual proteins from the T-Gel[™] Adsorbent; however, to avoid the possibility of cross-contaminating samples, dedicate each column for a particular application.

8. Place the bottom cap on the column, add 3 ml of storage buffer and apply the top cap to the column. Store column upright at 4°C.

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Related Pierce Products

37501	ImmunoPure [®] Monoclonal Antibody Isotyping Kit 1 (HRP/ABTS)
23310	Easy-Titer [®] Human IgG Assay Kit
23300	Easy-Titer [®] Mouse IgG Assay Kit
23305	Easy-Titer [®] Rabbit IgG Assay Kit
44887	ImmunoPure [®] IgM Fragmentation Kit
53004	EZ-Label TM Fluorescein Isothiocyanate (FITC) Protein Labeling Kit
53002	EZ-Label [™] Rhodamine Protein Labeling Kit

Product References

Harsay, E. and Schekman, R. (2002). A subset of yeast vacuolar protein sorting mutants is blocked in one branch of the exocytic pathway. J. Cell Biol. 156(2):271-85.

Koustova, E. et al. (2001). LP-BM5 virus-infected mice produce activating autoantibodies to the AMPA receptor. J. Clin. Invest. 107(6):737-44.

Suh, J.S., *et al.* (1998). Antibodies from patients with heparin-induced thrombocytopenia/thrombosis recognize different epitopes on heparin: platelet factor 4. *Blood.* **91(3)**:916-22.

General References

Porath, J., et al. (1985). Thiophilic adsorption-a new method for protein fractionation. FEBS Lett. 185:306-10.

Hutchens, T.W. and Porath, J. (1986). Thiophilic adsorption of immunoglobulins—analysis of conditions optimal for selective immobilization and purification. *Anal. Biochem.* **159:**217-26.

Belew, M., et al. (1987). A one-step purification method for monoclonal antibodies based on salt-promoted adsorption chromatography on a 'thiophilic' adsorbent. J. Immunol. Meth. **102:**173-82.

Nopper, B., *et al.* (1989). A thiophilic adsorbent for the one-step high-performance liquid chromatography purification of monoclonal antibodies. *Anal. Biochem.* **180:**66-71.

Oscarsson, S., et al. (1991). Thiophilic adsorbents for RIA and ELISA procedures. J. Immunol. Meth. 143:143-9.

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