

# ***Strep*-tagged Protein Purification Handbook**

For expressing, purifying, and detecting proteins carrying a *Strep*-tag® II or a 6xHis tag and a *Strep*-tag II

Two-step protein purification system

His-*Strep* pQE-TriSystem Vector Set

pQE-TriSystem *Strep* Vector

*Strep*-Tactin® Superflow and Superflow Cartridges

*Strep*-Tactin Magnetic Beads

*Strep*-tag Antibody



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*Strep-tag* technology for protein purification and detection is covered by US patent 5,506,121, UK patent 2272698 and French patent 93 13 066 and *Strep-Tactin* is covered by US patent 6,103,493.

Hoffmann-La Roche owns patents and patent applications pertaining to the application of Ni-NTA resin (Patent series: RAN 4100/63; USP 4.877.830, USP 5.047.513, EP 253 303 B1), and to 6xHis-coding vectors and His-labeled proteins (Patent series: USP 5.284.933, USP 5.130.663, EP 282 042 B1). All purification of recombinant proteins by Ni-NTA chromatography for commercial purposes, and the commercial use of proteins so purified, require a license from Hoffmann-La Roche.

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## Kit Contents

<b><i>Strep-Tactin</i><sup>®</sup> Superflow Cartridge</b>		<b>30120</b>
<i>Strep-Tactin</i> Superflow Cartridge (1 ml)		1
Handbook		1
<b><i>Strep-Tactin</i> Superflow</b>	<b>30001</b>	<b>30003</b>
<i>Strep-Tactin</i> Superflow	2 ml	10 ml
Handbook	1	1
<b><i>Strep-Tactin</i> Magnetic Beads</b>	<b>36311</b>	<b>36315</b>
<i>Strep-Tactin</i> Magnetic Beads	2 x 1 ml	20 x 1 ml
Handbook	1	1
<b><i>Strep-tag</i> Antibody</b>		<b>34850</b>
Mouse monoclonal antibody that recognizes the <i>Strep-tag</i> II epitope; lyophilized, for 1000 ml working solution		100 µg
Product sheet		1
Handbook		1
<b>pQE-TriSystem <i>Strep</i> Vector</b>		<b>33913</b>
pQE-TriSystem <i>Strep</i> vector		25 µg
Handbook		1
<b>His-<i>Strep</i> pQE-TriSystem Vector Set</b>		<b>32942</b>
pQE-TriSystem His- <i>Strep</i> 1 and pQE-TriSystem His- <i>Strep</i> 2 vectors		25 µg each
Handbook		

## Storage and Stability

*Strep-Tactin* matrices should be stored at 2–8°C. Under these conditions, *Strep-Tactin* matrices can be stored for up to 6 months without any reduction in performance. *Strep-Tactin* matrices should not be frozen.

*Strep-tag* Antibodies should be stored lyophilized until they are ready to be used. They can be stored lyophilized for 6 months at 2–8°C. In solution they can be stored for 3 months at 2–8°C or for 6 months in aliquots at –20°C. Avoid repeated freezing and thawing.

pQE-TriSystem His-*Strep* Vectors are supplied lyophilized with sucrose and bromophenol blue for visualization, and should be resuspended in a convenient volume of TE buffer (e.g., 10 µl) and stored at –20°C. Sucrose and bromophenol blue do not interfere with restriction digestions or bacterial transformation.

## Product Use Limitations

The kits and reagents described in this handbook are intended for research use. No claim or representation is intended to provide information for the diagnosis, prevention, or treatment of a disease. All due care and attention should be exercised in the handling of the products. We recommend all users of QIAGEN products to adhere to the NIH guidelines that have been developed for recombinant DNA experiments, or to other applicable guidelines.

## Quality Control

In accordance with QIAGEN's ISO-certified Quality Management System, each lot of kits and reagents described in this handbook is tested against predetermined specifications to ensure consistent product quality.

## Technical Assistance

At QIAGEN we pride ourselves on the quality and availability of our technical support. Our Technical Service Departments are staffed by experienced scientists with extensive practical and theoretical expertise in molecular biology and the use of QIAGEN® products. If you have any questions or experience any difficulties regarding the products described in this handbook or QIAGEN products in general, please do not hesitate to contact us.

QIAGEN customers are a major source of information regarding advanced or specialized uses of our products. This information is helpful to other scientists as well as to the researchers at QIAGEN. We therefore encourage you to contact us if you have any suggestions about product performance or new applications and techniques.

For technical assistance and more information please call one of the QIAGEN Technical Service Departments or local distributors (see back cover).

## Product Warranty and Satisfaction Guarantee

QIAGEN guarantees the performance of all products in the manner described in our product literature. The purchaser must determine the suitability of the product for its particular use. Should any product fail to perform satisfactorily due to any reason other than misuse, QIAGEN will replace it free of charge or refund the purchase price. We reserve the right to change, alter, or modify any product to enhance its performance and design. If a QIAGEN product does not meet your expectations, simply call your local Technical Service Department or distributor. We will credit your account or exchange the product — as you wish. Separate conditions apply to QIAGEN scientific instruments, service products, and to products shipped on dry ice. Please inquire for more information.

A copy of QIAGEN terms and conditions can be obtained on request, and is also provided on the back of our invoices. If you have questions about product specifications or performance, please call QIAGEN Technical Services or your local distributor (see back cover).

## Safety Information

When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. For more information, please consult the appropriate material safety data sheets (MSDSs). These are available online in convenient and compact PDF format at [www.qiagen.com/ts/msds.asp](http://www.qiagen.com/ts/msds.asp) where you can find, view, and print the MSDS for each QIAGEN kit and kit component.

The following risk and safety phrases apply to *Strep*-tag Antibodies:

### ***Strep*-tag Antibodies**

Sensitizer. Risk and safety phrases\*: R42/43. S24-26-36/37

### **24-hour emergency information**

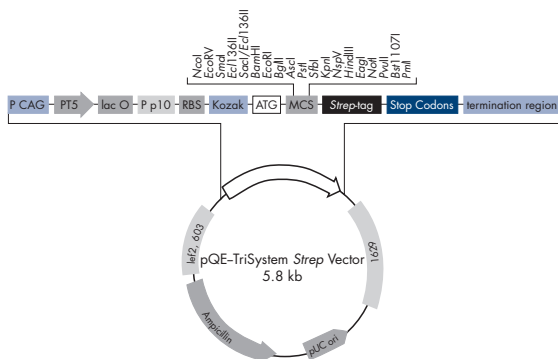
Emergency medical information in English, French, and German can be obtained 24 hours a day from:

Poison Information Center Mainz, Germany, Tel: +49-6131-1924

\* R42/43: May cause sensitization by inhalation and skin contact. S24: Avoid contact with skin; S26: In case of contact with eyes, rinse immediately with plenty of water and seek medical advice; S36/37: Wear suitable protective clothing and gloves.

# Introduction

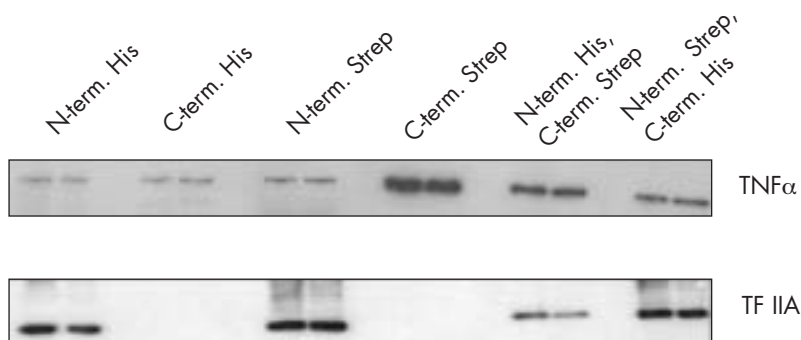
The *Strep*-tag II is a short peptide (8 amino acids, WSHPQFEK), which binds with high selectivity to *Strep*-Tactin, an engineered streptavidin. The *Strep*-tag II allows affinity chromatography on immobilized *Strep*-Tactin under physiological conditions, enabling native, active *Strep*-tagged proteins can be purified in a single step. The binding affinity of the *Strep*-tag II to *Strep*-Tactin ( $K_d = 1 \mu\text{M}$ ) is nearly 100 times higher than to streptavidin. After a short washing step, gentle elution of purified recombinant protein is performed by addition of low concentrations of biotin or desthiobiotin. Desthiobiotin is a stable, reversibly binding analog of biotin, the natural ligand of streptavidin. Proteins containing the *Strep*-tag II epitope can be detected with high specificity and sensitivity using *Strep*-tag Antibodies (Figure 3, page 8).



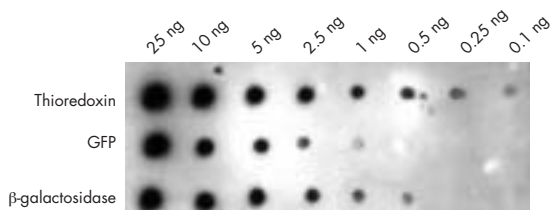
**Figure 1. pQE-TriSystem *Strep* vector for parallel protein expression using a single construct in *E. coli*, insect, and mammalian cells.** PT5: T5 promoter, lac O: lac operator, RBS: ribosome binding site, ATG: start codon, MCS: multiple cloning site, **Strep-tag**: *Strep*-tag sequence, **Stop Codons**: stop codons in all three reading frames, **Ampicillin**: ampicillin resistance gene, **P CAG**: CMV/actin/globin promoter, **P p10**: p10 promoter, **Kozak**: Kozak consensus sequence, **termination region**: transcription terminator region, **lef2, 603/1629**: flanking baculovirus sequences to permit generation of recombinant baculoviruses, **pUC ori**: pUC origin of replication.

## Construct screening for the best expression template

Because it contains just 8 amino acids (WSHPQFEK), the *Strep*-tag has a minimal effect on protein structure and function. However, expression-clone screening experiments have shown that addition of a *Strep*-tag to a recombinant protein construct can increase expression and solubility for specific proteins (Figure 2). Figure 2 also shows that the tag position can have an effect on expression; a tag can be required at the N-terminus to avoid secondary mRNA structures that inhibit initiation of translation. Both examples show that construct screening is a valuable tool for identifying the best expression template.



**Figure 2. Screening for the best expressing template can improve yields of soluble protein.** Six TF IIA  $\alpha\beta$  (TF IIA) and tumor necrosis factor  $\alpha$  (TNF $\alpha$ ) expression clone constructs with the indicated affinity tags were generated by PCR using the EasyXpress Linear Template Kit Plus and expressed in small-scale *in vitro* expression reactions using the EasyXpress Protein Synthesis Mini Kit. Expression level and ratio of soluble protein was investigated by separating equal volumes of total reaction (= total protein, left-hand lane in each pair) and supernatant after centrifugation (= soluble protein, right-hand lane in each pair) on an SDS-PAGE gel. Proteins were visualized using a mixture of *Strep*-tag and His-tag antibodies after western blotting.



**Figure 3. Dot blot showing the sensitivity of *Strep*-tag Antibody.** The indicated amounts of protein were spotted onto a membrane and detected using *Strep*-tag Antibody, an anti-mouse secondary antibody conjugated to horseradish peroxidase, and the ECL™ chemiluminescent detection system.

## Purification scale

Protocols are provided in this handbook for lysis of *E. coli*, insect, and mammalian cells. The purification scale is dependent on the amount of protein in the preparation. A column size and total binding capacity should be chosen to approximately match the amount of protein to be purified (see Table 1). Very few nontagged proteins will be retained on the resin when nearly all available binding sites are occupied by the tagged protein. If too much matrix is used, other proteins may bind nonspecifically to unoccupied sites and elute as contaminants.

High-yield preparations from *E. coli* cultures can be purified using a batch or FPLC® procedure. Eukaryotic expression systems typically deliver smaller amounts of protein than prokaryotic systems. This requires that smaller amounts of purification matrix be used for high-efficiency purification. Proteins that are obtained in high yields from baculovirus-infected insect cells can be purified using batch procedures. However, if protein expression is low, a purification procedure using magnetic beads is the method of choice. *Strep*-tagged proteins expressed in mammalian cells should be purified using *Strep*-Tactin Magnetic Beads. Table 1 lists the protein binding capacities of *Strep*-Tactin matrices.

**Table 1. Protein binding capacities of *Strep*-Tactin matrices**

Matrix	Protein binding capacity
<i>Strep</i> -Tactin Superflow	~1 mg/ml (~50 nmol @ ~20 kDa)
<i>Strep</i> -Tactin Magnetic Beads	200–300 µg/ml (10% suspension) (~10–15 nmol @ ~20 kDa)

## Buffer and reagent compatibility

Some reagents and buffer components used for protein purification may interfere with the affinity interaction between the protein and purification matrix. Table 2 lists compatibility of various reagents with the *Strep*-tag–*Strep*-Tactin interaction.

**Table 2. Reagents compatible with the *Strep*-tag–*Strep*-Tactin interaction\***

250 mM imidazole	5 mM DTT
1% Triton® X-100	1 M NaCl
1% Tween® 20	10% glycerol
0.3% CHAPS	1 mM EDTA
2% Igepal CA-630 (Nonidet P40)	

\* The reagents listed have been successfully used in concentrations up to those given.

## Protocol: Growth of *E. coli* Cultures (50 ml)

### Materials and equipment to be supplied by user\*

- LB medium
- Antibiotics
- IPTG

Buffer compositions are provided in Appendix B on page 41.

### Procedure

- 1. Inoculate 10 ml of LB medium containing the appropriate antibiotics in a 50 ml flask. Grow the cultures at 37°C overnight.**
- 2. Inoculate 50 ml of prewarmed media (with appropriate antibiotics) with 2.5 ml of the overnight cultures and grow at 37°C with vigorous shaking until an OD<sub>600</sub> of 0.6 is reached (30–60 min).**
- 3. Take a 0.5 ml sample immediately before induction.**  
This sample is the noninduced control; pellet cells and resuspend them in 25 µl 5x SDS-PAGE sample buffer. Freeze and store the sample at –20°C until SDS-PAGE analysis.
- 4. Induce expression by adding IPTG to a final concentration of 1 mM.**
- 5. Incubate the cultures for an additional 4–5 h. Collect a second 0.5 ml sample.**  
This sample is the induced control; pellet cells and resuspend them in 50 µl 5x SDS-PAGE sample buffer. Freeze and store the sample at –20°C until SDS-PAGE analysis.
- 6. Harvest the cells by centrifugation at 4000 x g for 20 min.**
- 7. Freeze and store cell pellet overnight at –20°C.**

\* When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. For more information, consult the appropriate material safety data sheets (MSDSs), available from the product supplier.

## Protocol: Preparation of Cleared Lysates from *E. coli* Cell Cultures

This lysis protocol uses *Strep*-Tactin Superflow Lysis Buffer and is suitable for subsequent single-step purification using a *Strep*-Tactin matrix.

For ultrahigh-purity preparations we recommend a two-step purification procedure using Ni-NTA Superflow followed by further purification using *Strep*-Tactin Superflow (see Appendix A). In such cases, use Ni-NTA Superflow Lysis Buffer for cell lysis.

The amount of cells required depends on the expression level of the *Strep*-tagged protein. For proteins that are expressed at high levels (10–50 mg of tagged protein per liter of cell culture), a 10x concentrated cell lysate (resuspend the pellet from a 50 ml culture in 5 ml lysis buffer) can be used.

Four ml of a 10x concentrated cell lysate in lysis buffer will contain approximately 0.4–2 mg of tagged protein.

For much lower expression levels (1–5 mg/liter), 200 ml of cell culture should be used to obtain a 50x concentrated cell lysate (4 ml cell lysate = 0.2–1 mg of tagged protein).

### Materials and equipment to be supplied by user\*

- 50 or 200 ml culture cell pellet (see note above)
- *Strep*-Tactin Superflow Lysis Buffer (Buffer NP, see note above)
- Lysozyme
- Optional: Benzonase® Nuclease

Buffer compositions are provided in Appendix B on page 41.

### Procedure

1. **Thaw the cell pellet for 15 min on ice and resuspend the cells in 5 ml lysis buffer.**
2. **Add lysozyme to 1 mg/ml and incubate on ice for 30 min.**
3. **Sonicate on ice.**

Use six 10 s bursts at 200–300 W with a 10 s cooling period between each burst. Use a sonicator equipped with a microtip.

4. **(Recommended) If the lysate is very viscous, add Benzonase® Nuclease (3 U per ml culture volume) and incubate on ice for 10–15 min.**

Alternatively, draw the lysate through a blunt-ended, narrow-gauge syringe needle several times.

\* When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. For more information, consult the appropriate material safety data sheets (MSDSs), available from the product supplier.

5. Centrifuge lysate at 10,000 x g for 20–30 min at 4°C to pellet the cellular debris. Save supernatant.

A certain proportion of the cellular protein may remain insoluble and will be located in the pellet.

6. Add 5  $\mu$ l 2x SDS-PAGE sample buffer to a 5  $\mu$ l aliquot of the supernatant and store at –20°C for SDS-PAGE analysis.
7. Proceed using one of the purification protocols.

See Table 3 to find the protocol best suited to your preparation scale.

**Table 3. Protocols for purification of His-*Strep*-tagged proteins from *E. coli* cell lysates**

	Expected amount of protein		
	$\leq 50 \mu\text{g}$	$> 50 \mu\text{g}$	$\gg 50 \mu\text{g}$
<b>Single-step purification</b>	Magnetic bead protocol, page 21	Superflow batch protocol, page 20	FPLC protocol, page 18
<b>Two-step purification</b>			
<b>Ni-NTA</b>	Magnetic bead protocol, page 39	Superflow batch protocol, page 38	FPLC protocol, page 37
<b><i>Strep</i>-Tactin</b>	Magnetic bead protocol, page 21	Superflow batch protocol, page 20	FPLC protocol, page 18

## Protocol: Preparation of Cleared Lysates from Baculovirus-Infected Insect Cells

Since they provide posttranslational modifications, insect cells are often the eukaryotic expression system of choice. Although expression rates are normally significantly higher in insect cells than in mammalian cells, expressed-protein levels are typically lower than those obtained in bacterial systems. The estimated total protein content in insect cells is approximately 20 mg per  $10^7$  cells. With recombinant protein expression levels ranging between 0.05% and 50%, protein yields are typically 10  $\mu$ g – 10 mg per  $10^7$  cells.

**Note:** If the expected yield of the preparation is less than 50  $\mu$ g, it is highly recommended that proteins be purified using the protocols for *Strep*-Tactin Magnetic Beads (page 21). Larger amounts of protein can be purified using a batch procedure. Lysis Buffer supplemented with 1% Igepal® CA-630 (Nonidet P40) is used for cell lysis. This lysis protocol uses *Strep*-Tactin Superflow Lysis Buffer and is suitable for subsequent single-step purification using a *Strep*-Tactin matrix.

For ultrahigh-purity preparations we recommend a two-step purification procedure using Ni-NTA Superflow followed by further purification using *Strep*-Tactin Superflow (see Appendix A). In such cases, use Ni-NTA Superflow Lysis Buffer for cell lysis.

### Materials and equipment to be supplied by user\*

- Cell pellet
- PBS
- *Strep*-Tactin Superflow Lysis Buffer (Buffer NP) supplemented with 1% Igepal® CA-630
- Optional: Benzonase® Nuclease

Buffer compositions are provided in Appendix B on page 41.

### Procedure

1. Wash the transfected cells with phosphate buffered saline (PBS) and collect them by centrifugation for 5 min at 1000 x g.
2. Lyse the cells in lysis buffer supplemented with 1% Igepal CA-630 by incubating for 10 min on ice. Use 4 ml lysis buffer per 1–2 x  $10^7$  cells.
3. (Recommended) If the lysate is very viscous, add Benzonase® Nuclease (3 U per ml culture volume) and incubate on ice for 10–15 min.

Alternatively, draw the lysate through a blunt-ended, narrow-gauge syringe needle several times.

\* When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. For more information, consult the appropriate material safety data sheets (MSDSs), available from the product supplier.

4. **Centrifuge the lysate at 10,000 x g for 10 min at 4°C to pellet cellular debris and DNA. Save the cleared lysate (supernatant).**

The supernatant should contain the tagged protein.

5. **Proceed using one of the purification protocols (see note above).**

# Protocol: Preparation of Lysates from Transfected Mammalian Cells

Recombinant proteins are often expressed in mammalian cells to allow eukaryotic posttranslational processing. However, expression levels are typically lower than in bacterial systems and often only small amounts of cell material are available. The total protein content in HeLa cells, for example, is around 3000  $\mu\text{g}$  per  $10^7$  cells. With recombinant protein expression levels at 0.01 to 1% of total protein, typical protein yields are 0.3 to 30  $\mu\text{g}$  per  $10^7$  cells. We therefore strongly recommend that proteins expressed in mammalian cells be purified using *Strep-Tactin* Magnetic Beads. Use of magnetic beads allows easy adjustment of binding capacity, and the small elution volumes provide concentrated proteins that are easily detected on SDS-PAGE gels.

This lysis protocol uses *Strep-Tactin* Beads Lysis Buffer (Buffer NP-T) and is suitable for subsequent single-step purification using *Strep-Tactin* Magnetic Beads.

For ultrahigh-purity preparations we recommend a two-step purification procedure using Ni-NTA Magnetic Agarose Beads followed by further purification using *Strep-Tactin* Magnetic Beads (see Appendix A). In such cases, use Ni-NTA Beads Lysis Buffer for cell lysis.

## Materials and equipment to be supplied by user\*

- Cell pellet
- PBS
- *Strep-Tactin* Beads Lysis Buffer (Buffer NP-T, see note above)
- Optional: Benzonase® Nuclease

Buffer compositions are provided in Appendix B on page 41.

## Procedure

1. **Wash the transfected cells with phosphate-buffered saline (PBS) and collect them by centrifugation for 5 min at 1000 x g.**
2. **Resuspend the cells in lysis buffer (containing 0.05% Tween 20) using 500  $\mu\text{l}$  lysis buffer per  $10^7$  cells.**

If higher concentrations of non-ionic detergent are required to solubilize the protein, detergents can be used in the lysis, wash, and elution buffers. Compatible detergents are Tween 20, Triton X-100, Igepal CA-630, and CHAPS (See Tables 2 and 5 on pages 9 and 34 respectively).

\* When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. For more information, consult the appropriate material safety data sheets (MSDSs), available from the product supplier.

**3. Lyse the cells according to steps 3a, 3b, or 3c.**

**3a. Lyse the cells by sonication on ice.**

Use six 15 s bursts at 75 W with a 10 s cooling period between each burst. Use a sonicator equipped with a microtip.

**3b. Lyse cells by three consecutive freeze–thaw cycles with freezing on dry ice and thawing at room temperature (15–25°C).**

**3c. With 0.5–1% non-ionic detergent (e.g., Tween 20 or Triton X-100) in the lysis buffer for solubilization, it is sufficient to incubate on an end-over-end shaker for 10 min at 4°C. Additional sonication or freeze–thaw cycles are not necessary.**

**4. (Recommended) If the lysate is very viscous, add Benzonase® Nuclease (3 U per ml culture volume) and incubate on ice for 10–15 min.**

Alternatively, draw the lysate through a blunt-ended, narrow-gauge syringe needle several times.

**5. Centrifuge the lysate at 10,000 x g for 10 min at 4°C to pellet cellular debris and DNA. Save the supernatant.**

The supernatant should contain the *Strep*-tagged protein.

**6. Proceed to protocol for purification using *Strep*-Tactin Magnetic Beads (page 21).**

# Protocol: Manual Purification of *Strep*-tagged Proteins Using a Syringe

This protocol is suitable for purifying *Strep*-tagged proteins using *Strep*-Tactin Superflow Cartridges and a syringe. The composition of the lysis and elution buffers can be modified to suit the particular application, e.g., by adding 0.1% Tween, 5 mM DTT, or 1 mM PMSF, or increasing NaCl or glycerol concentrations. See Table 2 on page 9 for a list of buffer reagents compatible with the *Strep*-tag–*Strep*-Tactin interaction.

## Things to do before starting

- Immediately before applying to column, sterile filter (0.2 or 0.45  $\mu\text{m}$ ) or centrifuge the lysate (10,000  $\times g$ ) to ensure that it is particle-free.

## Materials and equipment to be supplied by user

- Cleared cell lysate containing *Strep*-tagged protein
- *Strep*-Tactin Superflow Lysis Buffer (Buffer NP) and *Strep*-Tactin Superflow Elution Buffer (Buffer NPD). All buffers should be sterile filtered (0.2 or 0.45  $\mu\text{m}$ ) before use.
- 10 ml syringes
- Connector adapter

Buffer compositions are provided in Appendix B on page 41.

## Procedure

1. **Fill syringe with *Strep*-Tactin Superflow Lysis Buffer, attach a suitable adapter, and expel air by depressing syringe plunger until buffer drips from the end of the adapter.**
2. **Attach syringe to the cartridge inlet and remove cartridge outlet stopper.**
3. **Equilibrate the cartridge with 10 column volumes of buffer.**  
Do not use high pressure to force buffer through the cartridge. Ideal flow rates are 1 ml/min (1 ml cartridges).
4. **Remove syringe and fill with cleared lysate. Apply cleared lysate to the cartridge using the same flow rate as in step 3.**
5. **Using a fresh syringe and the same flow rate as in steps 3 and 4, wash the cartridge with 10 column volumes of *Strep*-Tactin Superflow Lysis Buffer.**
6. **Fill a syringe with *Strep*-Tactin Superflow Elution Buffer and elute protein from cartridge using the recommended flow rate.**

Protein usually elutes within 5–10 column volumes. *Strep*-Tactin Superflow Cartridges can be regenerated using the procedure provided in Appendix C on page 45.

# Protocol: Purification of *Strep*-tagged Proteins Using Liquid Chromatography Systems

This protocol is suitable for purifying *Strep*-tagged proteins using *Strep*-Tactin Superflow Cartridges on liquid chromatography systems (such as the ÄKTAdesign™ or FPLC™ System). During equilibration, loading, washing, and elution, monitor the back pressure generated by the chromatography system. Do not allow back pressure to exceed 5 bar (0.5 MPa).

The composition of the lysis and elution buffers can be modified to suit the particular application, e.g., by adding 0.1% Tween, 5 mM DTT, or 1 mM PMSF, or increasing NaCl or glycerol concentrations. See Table 2 on page 9 for a list of buffer reagents compatible with the *Strep*-tag–*Strep*-Tactin interaction.

## Materials and equipment to be supplied by user\*

- Cleared cell lysate containing *Strep*-tagged protein. Immediately before applying it to the column, sterile filter (0.2 or 0.45 µm) or centrifuge the lysate (10,000 x g) to ensure that it is particle-free.
- *Strep*-Tactin Superflow Lysis Buffer (Buffer NP) and *Strep*-Tactin Superflow Elution Buffer (Buffer NPD). All buffers should be sterile filtered (0.2 or 0.45 µm) before use.
- Connector adapter(s)

Buffer compositions are provided in Appendix B on page 41.

## Procedure

1. **Fill system pumps with *Strep*-Tactin Superflow Lysis Buffer and attach cartridge to the pump outlet, taking care not to introduce air into the system.**
2. **Remove cartridge outlet stopper and attach to the system tubing.**
3. **Equilibrate the cartridge with 10 column volumes of buffer.**  
Use a flow rate of 1 ml/min (1 ml cartridges).
4. **Using the system pumps or a superloop load the cleared cell lysate onto the cartridge.**

To ensure that the target protein has bound to the *Strep*-Tactin resin, retain the flow-through fraction for analysis by SDS-PAGE.

\* When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. For more information, consult the appropriate material safety data sheets (MSDSs), available from the product supplier.

5. **Fill system pumps with *Strep*-Tactin Superflow Lysis Buffer and wash the cartridge until the  $A_{280}$  returns to the baseline value.**

Retain the wash fraction for analysis by SDS-PAGE.

6. **Fill system pumps with *Strep*-Tactin Superflow Elution Buffer and elute protein from cartridge.**

Protein usually elutes within 5–10 column volumes. A shallow linear gradient over 20 column volumes may help separate proteins. *Strep*-Tactin Superflow Cartridges can be regenerated using the procedure provided in Appendix C on page 45.

## Protocol: Batch Purification of *Strep*-tagged Proteins Using *Strep*-Tactin Superflow

The composition of the lysis and elution buffers can be modified to suit the particular application, e.g., by adding 0.1% Tween, 5 mM DTT, or 1 mM PMSF, or increasing NaCl or glycerol concentrations. See Table 2 on page 9 for a list of buffer reagents compatible with the *Strep*-tag–*Strep*-Tactin interaction.

### Materials and equipment to be supplied by user

- Cleared lysate
- *Strep*-Tactin Superflow Lysis Buffer (Buffer NP)
- Empty columns
- *Strep*-Tactin Superflow Elution Buffer (Buffer NPD)
- *Strep*-Tactin Superflow

Buffer compositions are provided in Appendix B on page 41.

### Procedure

1. **Pipet cleared cell lysate into a 15 ml tube and add 2 ml *Strep*-Tactin Superflow Resin suspension. Mix gently by shaking (200 rpm on a rotary shaker) at 4°C for 60 min.**
2. **Load the *Strep*-Tactin Superflow resin into a column with the bottom outlet capped.**
3. **Remove bottom cap and collect the flow-through.**  
Save flow-through for SDS-PAGE analysis.
4. **Wash column two times with 4 ml *Strep*-Tactin Superflow Lysis Buffer.**  
Collect wash fractions for SDS-PAGE analysis.
5. **Elute the protein six times with 0.5 ml *Strep*-Tactin Superflow Elution Buffer. Collect eluates in six tubes and analyze by SDS-PAGE.**  
*Strep*-Tactin Superflow can be regenerated using the procedure provided in Appendix C on page 45.

## Protocol: Micro-Scale Purification of *Strep*-tagged Proteins Using *Strep*-Tactin Magnetic Beads

The composition of the lysis and elution buffers can be modified to suit the particular application, for example by adding 0.1% Tween, 5 mM DTT, or 1 mM PMSF, or increasing NaCl or glycerol concentrations. See Table 2 on page 9 for a list of buffer reagents compatible with the *Strep*-tag–*Strep*-Tactin interaction.

The amount of tagged protein recovered using *Strep*-Tactin Magnetic Beads is proportional to the protein concentration in the sample. Therefore, if samples contain low concentrations of protein, we recommended removing the storage buffer from the *Strep*-Tactin Magnetic Beads before adding the sample to the beads.

### Materials and equipment to be supplied by user\*

- Cleared lysate
- *Strep*-Tactin Magnetic Beads
- *Strep*-Tactin Beads Lysis Buffer
- *Strep*-Tactin Beads Elution Buffer
- Optional: Benzonase® Nuclease

Buffer compositions are provided in Appendix B on page 41.

### Procedure

1. Resuspend *Strep*-Tactin Magnetic Beads by vortexing for 2 s and then immediately add 200  $\mu$ l of 10% *Strep*-Tactin Magnetic Bead suspension to a cleared lysate or the pooled fractions eluted from Ni-NTA Magnetic Beads.

**Note:** Care is necessary to ensure that constant amounts of beads are pipetted. The beads will settle if the suspension is not agitated regularly. 200  $\mu$ l of *Strep*-Tactin Magnetic Beads suspension has a binding capacity of 40–60  $\mu$ g protein (see Table 1, page 9). If significantly different amounts of tagged protein are present in your lysate, the volume of magnetic-bead suspension should be varied accordingly. However, use of volumes less than 10  $\mu$ l are not recommended due to the associated handling problems — smaller volumes are difficult to pipet and may lead to uneven distribution of beads and reduced reproducibility. Generally, we recommend using cleared lysates for binding to *Strep*-Tactin Magnetic Beads. However, it may be possible to obtain good results by using crude lysates without clearing them. In this case, add Benzonase® Nuclease (3 U per ml culture volume), and incubate on ice for 10–15 min.

\* When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. For more information, consult the appropriate material safety data sheets (MSDSs), available from the product supplier.

2. **Mix the suspension gently on an end-over-end shaker for 30–60 min at 4°C.**
3. **Place the tube on a magnetic separator for 1 min and remove supernatant with a pipet.**

Tubes may be briefly centrifuged, before placing on the magnetic separator, to collect droplets of suspension from the tube caps.

4. **Remove the tube from the magnet, add 500  $\mu$ l *Strep*-Tactin Beads Lysis Buffer, gently vortex the suspension, place the tube on a magnetic separator for 1 min, and remove buffer.**

5. **Repeat step 4.**

Remaining buffer should be removed completely.

6. **Add 50  $\mu$ l *Strep*-Tactin Beads Elution Buffer, gently vortex the suspension, incubate the tube for 5 min, place the tube on a magnetic separator for 1 min, and collect the eluate in a clean tube.**

Tubes may be centrifuged before placing on the magnetic separator, to collect droplets of suspension from the tube caps.

7. **Repeat step 6 three times to give four eluate fractions.**

Because the biotin used for elution binds *Strep*-Tactin with extremely high affinity, it is not possible to regenerate *Strep*-Tactin Magnetic Beads.

## Protocol: Immunodetection using the *Strep*-tag Antibody (Chemiluminescent Method)

This protocol is used for chemiluminescent detection of *Strep*-tagged proteins on western or dot blots.

### Materials and equipment to be supplied by user\*

- Western blot or dot blot
- TBS (10 mM Tris-Cl; 150 mM NaCl, pH 7.5)
- TBS-Tween/Triton (20 mM Tris-Cl; 500 mM NaCl; 0.05% (v/v) Tween; 0.2% (v/v) Triton, pH 7.5)
- Blocking buffer (3% BSA or 1% casein in TBS)
- *Strep*-tag Antibody stock solution: Dissolve the lyophilized *Strep*-tag Antibody in 500  $\mu$ l water. The reconstituted solution contains 200  $\mu$ g/ml *Strep*-tag Antibody in PBS with PEG, sucrose, and sodium azide (0.08% [w/v]).
- Anti-mouse secondary antibody conjugate
- Secondary antibody dilution buffer (10% milk powder or 1% casein in TBS)

For compositions and preparation of buffers and reagents, see Appendix B, page 41.

The solutions required depend on the detection method employed.

For chemiluminescent detection, BSA does not sufficiently block nonspecific binding of the secondary antibody to the membrane, and milk powder should be used to dilute the secondary antibody. Alternatively, if alkali-soluble casein (Merck, Cat. No. 1.02241) is available in your country it can be used as a blocking reagent throughout the chemiluminescent detection protocol. The reagents used are shown in Table 4.

**Table 4. Reagents used in chemiluminescent detection of *Strep*-tagged proteins**

Step		(Alternative method)
Blocking	3% BSA in TBS	1% casein in TBS
<i>Strep</i> -tag Antibody binding	3% BSA in TBS	1% casein in TBS
Secondary antibody binding	10% milk powder in TBS	1% casein in TBS

### Chemiluminescent substrates

CDP-Star™ from Tropix, Inc. can be used with AP-conjugated secondary antibodies, and the ECL system from Amersham Pharmacia Biotech can be used in combination with HRP-conjugated secondary antibodies.

\* When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. For more information, consult the appropriate material safety data sheets (MSDSs), available from the product supplier.

## Procedure

- 1. Wash membrane twice for 10 min each time with TBS buffer at room temperature (15–25°C).**
- 2. Incubate for 1 h in blocking buffer at room temperature.**  
3% BSA (w/v) in TBS buffer\*, is used for blocking until incubation.
- 3. Wash membrane twice for 10 min each time in TBS-Tween/Triton buffer at room temperature.**
- 4. Wash membrane for 10 min with TBS buffer at room temperature.**
- 5. Incubate with *Strep*-tag Antibody (1/1000–1/2000 dilution of antibody stock solution in blocking buffer) at room temperature for 1 h.**  
Membrane can be sealed in plastic bags. 3% BSA (w/v) in TBS buffer\* is used for this blocking step when using chemiluminescent detection.
- 6. Wash twice for 10 min each time in TBS-Tween/Triton buffer at room temperature.**
- 7. Wash for 10 min in TBS buffer at room temperature.**
- 8. Incubate with secondary antibody solution for 1 h at room temperature.**  
Either alkaline phosphatase (AP) or horseradish peroxidase (HRP) conjugated anti-mouse IgG may be used. Dilute according to the manufacturer's recommendations. Use the lowest recommended concentration to avoid false signals.  
10% nonfat dried milk in TBS\* is used for incubation with secondary antibody when using chemiluminescent detection.  
Milk powder is needed to reduce background because BSA does not block sufficiently for the very sensitive chemiluminescent detection method.
- 9. Wash 4 times for 10 min each time in TBS-Tween/Triton buffer at room temperature.**
- 10. Perform chemiluminescent detection reaction and expose to X-ray film according to the manufacturer's recommendations.**

\* If alkali-soluble casein (Merck, Cat. No. 1.02241) is available in your country a 1% (w/v) solution in TBS buffer can be used for this protocol step.

# Protocol: Immunodetection using the *Strep*-tag Antibody (Chromogenic Method)

The solutions required depend on the detection method employed.

## Materials and equipment to be supplied by user\*

- Western Blot, dot blot, or colony blot
- TBS (10 mM Tris-Cl; 150 mM NaCl, pH 7.5)
- TBS-Tween/Triton (20 mM Tris-Cl; 500 mM NaCl; 0.05% [v/v] Tween; 0.2% [v/v] Triton, pH 7.5)
- *Strep*-tag Antibody
- Anti-mouse secondary antibody alkaline phosphatase (AP) or horseradish peroxidase (HRP) conjugate
- Blocking buffer (3% BSA in TBS)
- Secondary antibody dilution buffer (3% BSA in TBS)
- Staining solutions for alkaline phosphatase (AP) or horseradish peroxidase (HRP)

The solutions required depend on the antibody and detection method used. For the chromogenic detection method, 3% (w/v) BSA in TBS is used as a blocking reagent throughout the whole procedure.

## Procedure

1. **Wash membrane twice for 10 min each time with TBS buffer at room temperature (15–25°C).**
2. **Incubate for 1 h in blocking buffer at room temperature.**  
3% BSA (w/v) in TBS buffer is used for blocking throughout the procedure when using chromogenic detection.
3. **Wash membrane twice for 10 min each time in TBS-Tween/Triton buffer at room temperature.**  
Use of TBS-Tween/Triton buffer has been found empirically to result in optimal signal-to-noise ratios.
4. **Wash membrane for 10 min with TBS buffer at room temperature.**
5. **Incubate with *Strep*-tag Antibody (1/1000–1/2000 dilution of antibody stock solution in blocking buffer) at room temperature for 1 h.**  
Membrane can be sealed in plastic bags.  
3% BSA (w/v) in TBS buffer, is used for blocking throughout the procedure when using chromogenic detection.

\* When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. For more information, consult the appropriate material safety data sheets (MSDSs), available from the product supplier.

6. Wash twice for 10 min each time in TBS-Tween/Triton buffer at room temperature.
7. Wash for 10 min in TBS buffer at room temperature.
8. Incubate with secondary antibody solution diluted in 3% BSA (w/v) in TBS for 1h at room temperature.

Either alkaline phosphatase (AP) or horseradish peroxidase (HRP) conjugated anti-mouse IgG may be used. Rabbit-anti-mouse IgG/AP-conjugate from Pierce (Cat. No. 31332) or goat-anti-mouse IgG/HRP-conjugate from Jackson ImmunoResearch (Cat. No. 115-035-003) yield good results. Dilute according to the manufacturer's recommendations. Use the lowest recommended amounts to avoid false signals.

9. Wash 4 times for 10 min each time in TBS-Tween/Triton buffer at room temperature.

#### Detection

10. Stain with AP or HRP staining solution until the signal is clearly visible (approximately 5–15 min).

Do not shake blots during color development.

11. Stop the chromogenic reaction by rinsing the membrane twice with water.
12. Dry the membrane and photograph as soon as possible.

The colors will fade with time. The product formed when using HRP is particularly unstable.

# Troubleshooting Guide

This troubleshooting guide may be helpful in solving any problems that may arise. The scientists in QIAGEN Technical Services are always happy to answer any questions you may have about either the information and protocols in this handbook or molecular biology applications (see back cover for contact information).

## Comments and suggestions

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### Purification using *Strep-Tactin* matrices

#### Protein does not bind to the *Strep-Tactin* matrix

- |  |  |
|--|--|
| a) Binding conditions are incorrect                                | Avoid using chaotropic salts in buffers. In contrast to 6xHis-tag–Ni-NTA interaction even low (<500 mM) concentrations of urea will disturb the <i>Strep</i> -tag II– <i>Strep-Tactin</i> interaction.<br>Check buffer pH, which should be above 6.5.  |
| b) <i>Strep-Tactin</i> matrix binding sites are occupied/blocked   | Biotin irreversibly blocks the binding sites of <i>Strep-Tactin</i> and should be removed from protein samples by dialysis; alternatively avidin can be added to protein samples. Addition of avidin does not interfere with the <i>Strep</i> -tag II– <i>Strep-Tactin</i> interaction.  |
| c) <i>Strep</i> -tag is not present                                | Sequence ligation junctions to ensure that the reading frame is correct.<br>Check for possible internal translation starts (N-terminal tag) or premature termination sites (C-terminal tag).   |
| d) <i>Strep</i> -tag is inaccessible                               | Move tag to the opposite end of the protein.   |
| e) <i>Strep</i> -tag– <i>Strep-Tactin</i> interaction is very weak | Replace batch purification by applying the lysate to a prepared column.<br>Use more concentrated lysate or Ni-NTA eluate and remove <i>Strep-Tactin</i> Magnetic Beads storage buffer prior to use.  |
| f) Protein does not elute from <i>Strep-Tactin</i> resin           | The <i>Strep</i> -tag II– <i>Strep-Tactin</i> interaction has a $K_D$ value in the $\mu\text{M}$ range. Therefore, tight binding is most probably caused by unspecific binding to the Superflow or Magnetic Agarose Beads matrix.<br>Elution with 10 mM biotin is possible but all <i>Strep-Tactin</i> binding sites will be blocked irreversibly. |
| g) Protein has precipitated on the column                          | Add detergents to or reduce salt concentration in buffers. Do not use a pH near the pI of the protein. Increase or lower pH (pH must however, be above 6.5).   |

### Purification using Ni-NTA matrices

#### Protein does not bind to the Ni-NTA resin

- |   |   |
|---|---|
| a) 6xHis tag is not present                 | Sequence ligation junctions to ensure that the reading frame is correct.<br>Check for possible internal translation starts (N-terminal tag) or premature termination sites (C-terminal tag).  |
| b) 6xHis tag is inaccessible                | Move tag to the opposite end of the protein.  |
| c) 6xHis tag has been degraded              | Check that the 6xHis tag is not associated with a portion of the protein that is processed.   |
| d) Binding conditions incorrect             | Check pH and composition of all buffers and solutions.<br>Ensure that there are no chelating or reducing agents present, and that the concentration of imidazole is not too high.   |
| e) Protein elutes in the wash buffer        | Wash stringency is too high. Lower the concentration of imidazole or increase the pH slightly.  |
| f) 6xHis tag is partially hidden            | Reduce wash stringency.   |
| g) Buffer conditions incorrect              | Check pH and composition of wash buffer. Ensure that there are no chelating or reducing agents present.   |
| h) Protein precipitates during purification | Temperature is too low. Perform purification at room temperature.   |
| i) Protein forms aggregates                 | Try adding solubilization reagents such as 0.1% Triton X-100 or Tween-20, up to 20 mM $\beta$ -ME, up to 2 M NaCl, or stabilizing cofactors such as $Mg^{2+}$ . These may be necessary in all buffers to maintain protein solubility. |

#### Protein does not elute

- |  |   |
|--|---|
| a) Elution conditions are too mild (protein may be in an aggregate or multimer form) | Elute with a pH or imidazole step gradient to determine the optimal elution conditions. |
| b) Protein has precipitated on the column  | Perform binding and elution in batch format to avoid high local protein concentrations. |

## Comments and suggestions

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### Protein elutes with contaminants

- a) Binding and wash conditions not stringent enough  
Include 10–20 mM imidazole in the binding and wash buffers.
- b) Contaminants are associated with tagged protein.  
Add  $\beta$ -ME to a maximum of 20 mM or DTT to 10 mM to reduce disulfide bonds.  
Increase salt and/or detergent concentrations, or add ethanol/glycerol to wash buffer to disrupt nonspecific interactions.
- c) Contaminants are truncated forms of the tagged protein  
Check for possible internal translation starts (C-terminal tag) or premature termination sites (N-terminal tag).  
Prevent protein degradation during purification by working at 4°C or by including protease inhibitors.

### Discoloration of resin

- Nickel ions are removed or reduced  
Ensure that there are no chelating compounds (resin turns white in color) or high concentrations of reducing agents (resin turns brown in color) present in the buffers.

### Purification from mammalian cells

- a) No protein band in SDS-PAGE analysis of the eluate  
Expression is too low. Check the expression level by western blotting using an Anti-His Antibody or a protein-specific antibody. Alternatively, perform an immunoassay with Ni-NTA Magnetic Agarose Beads (see the *Ni-NTA Magnetic Agarose Beads Handbook*) or ELISA using Ni-NTA HisSorb Strips (see the *QIAexpress Detection and Assay Handbook*). If only small amounts of 6xHis-tagged protein are present in the lysate, increase the amount of starting cell material and purify with an equal amount of magnetic beads. Do not exceed lysis volumes of 2 ml — this allows purification in a single 2 ml tube.
- b) 6xHis tagged protein has been degraded  
Check that the 6xHis tag is not removed from the protein during post-translational processing.  
Work at 4°C and add protease inhibitors, such as PMSF.

## Comments and suggestions

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- c) 6xHis-tagged protein partially elutes in the wash
- The binding capacity used is too low to bind all of the 6xHis-tagged protein. 10  $\mu$ l magnetic-bead suspension has a binding capacity of 3  $\mu$ g 6xHis-tagged DHFR (24 kDa). If significantly larger amounts of 6xHis-tagged protein are present in the lysate, increase the amount of beads accordingly.

### Binding of contaminants

- a) Too much Ni-NTA matrix was used
- Match the total binding capacity of the beads to the amount of 6xHis-tagged protein to be purified by simply adjusting the amount of Ni-NTA Magnetic Agarose Beads suspension used.
- Proteins that contain neighboring histidines are not common in bacteria, but do occur in eukaryotic cells. These proteins, as well as endogenous proteins with metal-binding sites, normally bind with lower affinity to the Ni-NTA matrix than do 6xHis-tagged proteins. If the binding capacity of the amount of beads used greatly exceeds the amount of 6xHis-tagged protein to be purified, these proteins will bind to the Ni-NTA matrix to a considerably higher extent, and will be subsequently recovered in the eluate.
- b) Binding and wash conditions are not stringent enough
- Always include 10–20 mM imidazole in the binding buffer and 20 mM imidazole in the wash buffer.
- c) Large amount of nontagged proteins in the lysate when purifying from cells with a low expression rate
- Perform a second round of purification from the eluate after adjusting the imidazole concentration to 10–20 mM using binding buffer without imidazole. Significantly smaller amounts of background proteins in the binding step reduce the level of contaminants in the final preparation.

### Purification from insect cells

#### No protein band in SDS-PAGE analysis of the fractions

- a) No or low expression      Check the expression level by western blotting using an Anti-His Antibody or a protein-specific antibody. Alternatively perform an ELISA using Ni-NTA HisSorb Strips (see *QIAexpress Detection and Assay Handbook*). If low amounts of 6xHis-tagged protein are present in the lysate, increase the amount of starting cell material and purify with an equal amount of Ni-NTA matrix
- b) 6xHis-tagged protein has been degraded      Check that the 6xHis tag is not removed from the protein during post-translational processing or by endogenous proteases during the purification procedure.  
Work at 4°C and add protease inhibitors, such as PMSF.
- c) 6xHis-tagged protein partially elutes in the wash buffer or flow-through      The amount of matrix used is too low to bind all of the 6xHis-tagged protein. 100 µl Ni-NTA agarose has a binding capacity of 500–1000 µg 6xHis-tagged protein. Adjust the amount of matrix used for purification accordingly.

#### Contaminants bind to resin

- a) Too much Ni-NTA matrix was used      Match the total binding capacity of the matrix used to the amount of 6xHis-tagged protein to be purified. Endogenous proteins with metal-binding sites normally bind with lower affinity to the Ni-NTA matrix than do 6xHis-tagged proteins. If the binding capacity of the amount of matrix used greatly exceeds the amount of 6xHis-tagged protein to be purified, these proteins will bind to the Ni-NTA matrix to a considerably higher extent, and subsequently will be recovered in the eluate.
- b) Binding and wash conditions are not stringent enough      Always include 10–20 mM imidazole in the binding buffer and 20 mM imidazole in the wash buffer.

## Appendix A: Two-Step Purification Using a His-tag and a *Strep*-tag

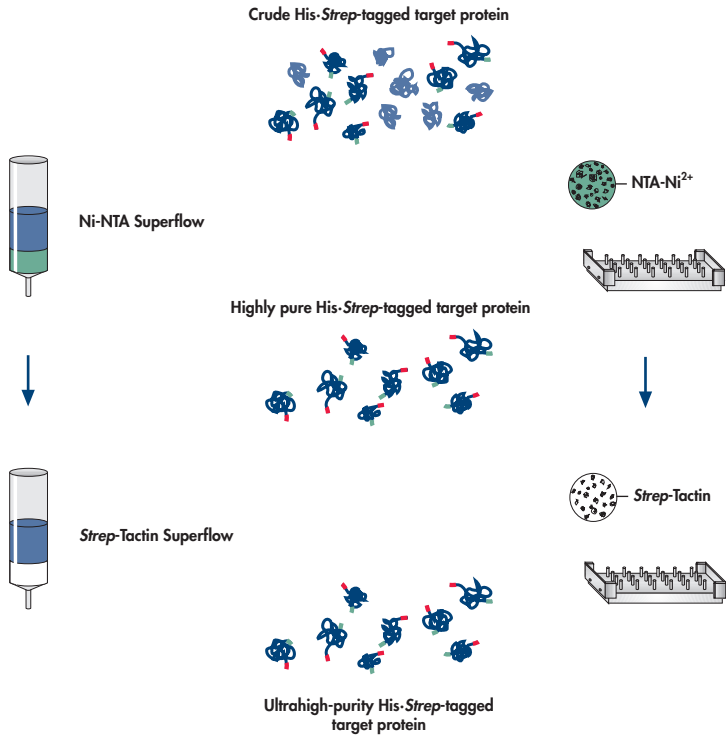
Many researchers in structural and functional proteomics use eukaryotic systems, such as insect or mammalian cells, to obtain protein posttranslational modifications not provided by conventional bacterial expression. However, the complexity of eukaryotic proteomes makes purification of proteins expressed using these systems a challenge and typically means that individual purification schemes must be developed for each protein. There is therefore an urgent need for a standardized method for purification of functional, high-purity proteins. In order to meet this need, QIAGEN offers a two-step affinity purification process, which provides ultrapure protein even from eukaryotic expression systems. To increase purity, proteins can be tagged with both a His- and *Strep*-tag and purified in a two-step procedure using Ni-NTA and *Strep*-Tactin. Recombinant proteins that carry the two small affinity tags are efficiently expressed in *E. coli*, insect, or mammalian cells using pQE-TriSystem His-*Strep* vectors (Figure 5, page 34). After cell lysis and clearing of the lysate, proteins are initially purified using an immobilized-metal affinity chromatography procedure that is based on the proven 6xHis-tag–Ni-NTA interaction. After elution from the Ni-NTA matrix using imidazole, recombinant proteins (which also carry the *Strep*-tag II epitope) are loaded directly onto a *Strep*-Tactin matrix (see flowchart, page 33). No buffer exchange is required. Protein is eluted from the *Strep*-Tactin matrix using either biotin or desthiobiotin. This two-step affinity purification delivers ultrapure (>98% pure) protein (Figure 4, page 33). The order of purifications can be reversed (i.e., *Strep*-Tactin followed by Ni-NTA purification).

### Initial purification of His-*Strep*-tagged proteins using the 6xHis-tag–Ni-NTA interaction

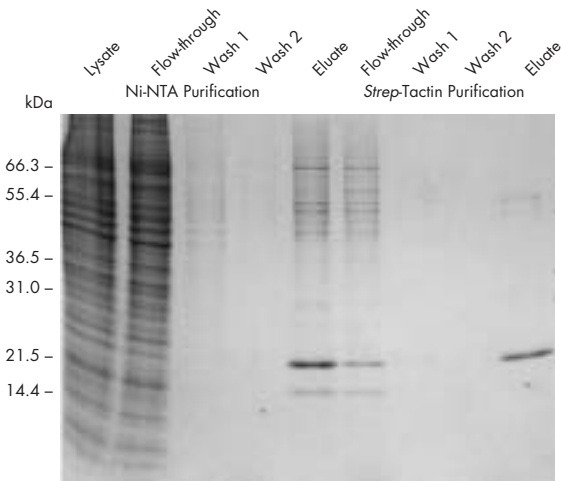
The initial stage of His-*Strep*-tagged protein purification is based on the remarkable selectivity and high affinity of patented Ni-NTA (nickel-nitrilotriacetic acid) resin for proteins containing an affinity tag of six consecutive histidine residues — the 6xHis tag. NTA, which has four chelation sites for nickel ions, binds nickel more tightly than metal-chelating purification systems that only have three sites available for interaction with metal ions. The extra chelation site prevents nickel-ion leaching; providing a greater binding capacity, and high-purity protein preparations.

**Table 5. Protein binding capacities of Ni-NTA matrices**

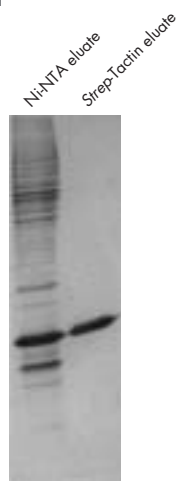
Matrix	Protein binding capacity
Ni-NTA Superflow and Agarose	5–10 mg/ml (250–500 nmol @ ~20 kDa)
Ni-NTA Magnetic Agarose Beads	300 µg/ml (5% suspension) (~15 nmol @ ~20 kDa)



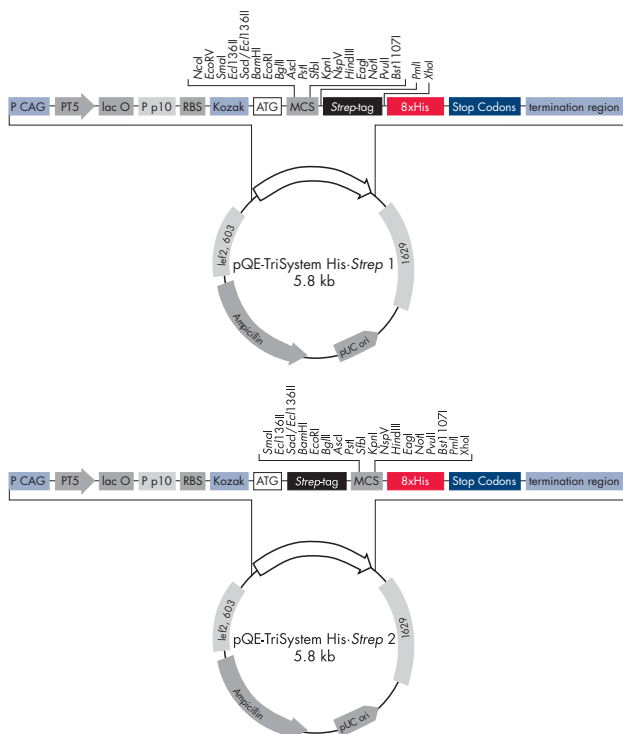
**A**



**B**



**Figure 4. Thioredoxin, expressed in NIH-3T3 cells, was purified using the Two-Step Affinity Purification System. A** Coomassie<sup>®</sup>-stained gel. **B** Silver-stained gel.



**Figure 5. pQE-TriSystem His-Strep vectors for parallel protein expression using a single construct in *E. coli*, insect, and mammalian cells.** PT5: T5 promoter, lac O: lac operator, RBS: ribosome binding site, ATG: start codon, 8xHis: His tag sequence, MCS: multiple cloning site, Strep-tag: Strep-tag sequence, Stop Codons: stop codons in all three reading frames, Ampicillin: ampicillin resistance gene, P CAG: CMV/actin/globin promoter, P p10: p10 promoter, Kozak: Kozak consensus sequence, termination region: transcription terminator region, lef2, 603/1629: flanking baculovirus sequences to permit generation of recombinant baculoviruses, pUC ori: pUC origin of replication.

Some reagents and buffer components used for protein purification may interfere with the affinity interaction between the protein and purification matrix. Table 5 lists compatibility of various reagents with the Ni-NTA–6xHis interaction.

**Table 5. Reagents compatible with the Ni-NTA–6xHis interaction\***

6 M guanidine HCl	50% glycerol	8 M urea
20% ethanol	2% Triton X-100	2 M NaCl
2% Tween 20	4 M MgCl <sub>2</sub>	1% CHAPS
5 mM CaCl <sub>2</sub>	20 mM β-mercaptoethanol	20 mM imidazole

\* The reagents listed have been successfully used in concentrations up to those given.

# Protocol: Protein Purification from Insect-Cell Growth Medium

To purify proteins secreted into insect-cell growth medium, the pH and salt concentrations of the medium must be adjusted to provide optimal conditions for protein binding to Ni-NTA. However, media used to culture insect cells may contain electron-donating groups (e.g., the amino acids glutamine, glycine, or histidine) that can prevent binding of 6xHis tagged proteins to Ni-NTA. If purification fails, dialysis, ultrafiltration, or gel filtration can be used to adjust the sample to the optimal buffer conditions for Ni-NTA purification.

## Materials and equipment to be supplied by user\*

- Insect-cell growth medium containing secreted proteins
- 10x Binding Buffer (500 mM NaH<sub>2</sub>PO<sub>4</sub>; 1.5 M NaCl, pH 8.0)

Buffer compositions are provided in Appendix B on page 41.

## Procedure

- 1. Add a volume of the 10x binding buffer to the insect-cell growth medium that will result in a 1x end concentration and mix.**

For instance, if you have 45 ml insect-cell medium, add 5 ml 10x binding buffer to give a total volume of 50 ml.

- 2. Proceed using one of the Ni-NTA purification protocols.**

\* When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. For more information, consult the appropriate material safety data sheets (MSDSs), available from the product supplier.

# Protocol: Manual Purification of His-tagged Proteins Using a Syringe

This protocol is for purification of His-tagged proteins using Ni-NTA Superflow Cartridges and a syringe. The composition of the lysis, wash, and elution buffers can be modified to suit the particular application, for example by adding 0.1% Tween, 5–10 mM  $\beta$ -mercaptoethanol, or 1 mM PMSF, or increasing NaCl or glycerol concentrations. See Table 5 on page 34 for a list of buffer reagents compatible with the 6xHis-tag–Ni-NTA interaction.

## Things to do before starting

- Immediately before applying to column, sterile filter (0.2 or 0.45  $\mu$ m) or centrifuge the lysate (10,000  $\times$  g) to ensure that it is particle-free.

## Materials and equipment to be supplied by user

- Cleared cell lysate containing 6xHis-tagged protein
- Buffers NPI-10, NPI-20, and NPI-250 (native conditions) or Buffers B, C, and E (denaturing conditions). All buffers should be sterile filtered (0.2 or 0.45  $\mu$ m) before use.
- 10 ml syringes
- Connector adapter

Buffer compositions are provided in Appendix B on page 41.

## Procedure

1. **Fill syringe with Buffer NPI-10 (native conditions) or Buffer B (denaturing conditions), attach a suitable adapter, and expel air by depressing syringe plunger until buffer drips from the end of the adapter.**
2. **Attach syringe to the cartridge inlet and remove cartridge outlet stopper.**
3. **Equilibrate the cartridge with 10 column volumes of buffer.**  
Do not use high pressure to force buffer through the cartridge. Ideal flow rates are 1 ml/min (1 ml cartridges) and 5 ml/min (5 ml cartridges).
4. **Remove syringe and fill with cleared lysate. Apply cleared lysate to the cartridge using the same flow rate as in step 3.**
5. **Using a fresh syringe and the same flow rate as in steps 3 and 4, wash the cartridge with 10 column volumes of Buffer NPI-20 (native conditions) or Buffer C (denaturing conditions).**
6. **Fill a syringe with Buffer-NPI 250 (native conditions) or Buffer E (denaturing conditions) and elute protein from cartridge using the recommended flow rate.**

Protein usually elutes within 5–10 column volumes.

## Protocol: Purification of His-tagged Proteins Using Liquid Chromatography Systems

This protocol is suitable for purifying His-tagged proteins using Ni-NTA Superflow Cartridges on liquid chromatography systems (such as the ÄKTA or FPLC System). During equilibration, loading, washing, and elution monitor the back pressure generated by the chromatography system. Do not allow back pressure to exceed 5 bar (0.5 MPa). The composition of the lysis, wash, and elution buffers can be modified to suit the particular application, for example by adding 0.1% Tween, 5–10 mM  $\beta$ -mercaptoethanol, or 1 mM PMSF, or increasing NaCl or glycerol concentrations. See Table 5 on page 34 for a list of buffer reagents compatible with the 6xHis-tag–Ni-NTA interaction.

### Materials and equipment to be supplied by user

- Cleared cell lysate containing 6xHis-tagged protein. Immediately before applying it to the column, sterile filter (0.2 or 0.45  $\mu$ m) or centrifuge the lysate (10,000  $\times$  g) to ensure that it is particle-free.
- Buffers NPI-10, NPI-20, and NPI-250 (native conditions) or Buffers B, C, and E (denaturing conditions). All buffers should be sterile filtered (0.2 or 0.45  $\mu$ m) before use
- Connector adapter(s)

Buffer compositions are provided in Appendix B on page 41.

### Procedure

1. **Fill system pumps with Buffer NPI-10 (native conditions) or Buffer B (denaturing conditions) and attach cartridge to the pump outlet, taking care not to introduce air into the system.**
2. **Remove cartridge outlet stopper and attach to the system tubing.**
3. **Equilibrate the cartridge with 10 column volumes of buffer.**

Use a flow rate of 1 ml/min (1 ml cartridges) or 5 ml/min (5 ml cartridges).

4. **Using the system pumps or a superloop load the cleared cell lysate onto the cartridge.**

To ensure that the target protein has bound to the Ni-NTA resin, retain the flow-through fraction for analysis by SDS-PAGE.

5. **Fill system pumps with Buffer NPI-20 (native conditions) or Buffer C (denaturing conditions) and wash the cartridge until the  $A_{280}$  returns to the baseline value.**

Retain the wash fraction for analysis by SDS-PAGE.

6. **Fill system pumps with Buffer-NPI 250 (native conditions) or Buffer E (denaturing conditions) and elute protein from cartridge.**

Protein usually elutes within 5–10 column volumes. A shallow linear gradient over 20 column volumes may help separate proteins.

## Protocol: Batch Purification of His-*Strep*-tagged Proteins Using Ni-NTA Matrices

Ni-NTA Resin Lysis Buffer contains 10 mM imidazole to minimize binding of untagged, contaminating proteins and increase purity with fewer wash steps. If the tagged protein does not bind under these conditions, the amount of imidazole should be reduced to 1–5 mM. With His-*Strep*-tagged proteins exhibiting high binding affinities, the imidazole concentration can be increased to 20 mM.

The composition of the lysis, wash, and elution buffers can be modified to suit the particular application, for example by adding 0.1% Tween, 5–10 mM  $\beta$ -mercaptoethanol, or 1 mM PMSF, or increasing NaCl or glycerol concentrations. See Table 5 on page 34 for a list of buffer reagents compatible with the 6xHis-tag–Ni-NTA interaction.

### Materials and equipment to be supplied by user\*

- Cleared cell lysate
- Buffers NPI-20 and NPI-250 (native conditions) or Buffers C and E (denaturing conditions). All buffers should be sterile filtered (0.2 or 0.45  $\mu$ m) before use.
- Ni-NTA matrix
- Empty columns

Buffer compositions are provided in Appendix B on page 41.

### Procedure

1. **Add 1 ml of 50% Ni-NTA matrix slurry to 4 ml cleared lysate and mix gently by shaking (200 rpm on a rotary shaker) at 4°C for 60 min.**
2. **Load the lysate–Ni-NTA mixture into a column with the bottom outlet capped.**
3. **Remove bottom cap and collect the column flow-through.**  
Save flow-through for SDS-PAGE analysis.
4. **Wash twice with 4 ml Buffer NPI-20 (native conditions) or Buffer C (denaturing conditions).**  
Collect wash fractions for SDS-PAGE analysis.
5. **Elute the protein four times with 0.5 ml Buffer-NPI 250 (native conditions) or Buffer E (denaturing conditions). Collect the eluate in four tubes and analyze by SDS-PAGE.**

\* When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. For more information, consult the appropriate material safety data sheets (MSDSs), available from the product supplier.

## Protocol: Micro-Scale Purification of His-*Strep*-tagged Proteins Using Ni-NTA Magnetic Agarose Beads

The composition of the lysis, wash, and elution buffers can be modified to suit the particular application, for example by adding 0.1% Tween, 5–10 mM  $\beta$ -mercaptoethanol, or 1 mM PMSF, or increasing NaCl or glycerol concentrations. See Table 5 on page 34 for a list of buffer reagents compatible with the 6xHis-tag–Ni-NTA interaction.

### Materials and equipment to be supplied by user\*

- Cleared lysate
- Ni-NTA Magnetic Agarose Beads
- Buffers NPI-20-T and NPI-250-T (native conditions) or Buffers C and E (denaturing conditions).
- Optional: Benzonase® Nuclease

Buffer compositions are provided in Appendix B on page 41.

### Procedure

1. **Resuspend the Ni-NTA Magnetic Agarose Beads by vortexing for 2 s and then immediately add 200  $\mu$ l of the 5% Ni-NTA Magnetic Agarose Bead suspension to each 1 ml of lysate containing the His-*Strep*-tagged protein.**

**Note:** Care is necessary to ensure that constant amounts of beads are pipetted. The beads will settle if the suspension is not agitated regularly. 200  $\mu$ l magnetic-bead suspension has a binding capacity of 60  $\mu$ g 6xHis-tagged DHFR (24 kDa). If significantly different amounts of tagged protein are present in your lysate, the volume of magnetic-bead suspension should be varied accordingly. However, use of volumes less than 10  $\mu$ l are not recommended due to the associated handling problems — smaller volumes are difficult to pipet and may lead to uneven distribution of beads and reduced reproducibility. Generally, we recommend using cleared lysates for binding to Ni-NTA Magnetic Agarose Beads. However, it may be possible to obtain good results by using crude lysates without clearing them. In this case, use lysates which have been diluted 5-fold, and if using native lysis conditions, add Benzonase® Nuclease (3 U per ml culture volume), and incubate on ice for 10–15 min.

\* When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. For more information, consult the appropriate material safety data sheets (MSDSs), available from the product supplier.

- 2. Mix the suspension gently on an end-over-end shaker for 30–60 min at room temperature (15–25°C).**

The time and temperature necessary for efficient binding is dependent on the protein and the accessibility of the 6xHis tag in the buffer system used. Especially under native conditions, it may be necessary to incubate at 4°C if the protein is not stable at room temperature.

- 3. Place the tube on a magnetic separator for 1 min and remove supernatant with a pipet.**

Tubes may be briefly centrifuged, before placing on the magnetic separator, to collect droplets of suspension from the tube caps.

- 4. Remove tube from the magnet, add 500 µl of Buffer NPI-20-T (native conditions) or Buffer C (denaturing conditions), mix the suspension, place the tube on a magnetic separator for 1 min, and remove buffer.**

- 5. Repeat step 4.**

Remaining buffer should be removed completely.

- 6. Add 100 µl of Buffer NPI-250-T (native conditions) or Buffer E (denaturing conditions), mix the suspension, incubate the tube for 1 min, place for 1 min on a magnetic separator, and collect the eluate in a clean tube.**

Tubes may be centrifuged before placing on the magnetic separator, to collect droplets of suspension from the tube caps.

- 7. Repeat step 6.**

Most of the 6xHis-tagged protein will elute in the first elution step. If a more concentrated protein solution is required, elute in two aliquots of 50 µl.

## Appendix B: Composition of Buffers

When working with chemicals, always wear a suitable lab coat, disposable gloves, and protective goggles. For more information, consult the appropriate material safety data sheets (MSDSs), available from the product supplier.

### LB medium

10 g/liter tryptone; 5 g/liter yeast extract; 10 g/liter NaCl

### 2x SDS-PAGE sample buffer

0.09 M Tris-Cl; 20% glycerol; 2% SDS; 0.02% bromophenol blue;  
0.1 M DTT, pH 6.8

### 5x SDS-PAGE sample buffer

0.225 M Tris-Cl; 50% glycerol; 5% SDS; 0.05% bromophenol blue;  
0.25 M DTT, pH 6.8

### 10x Binding Buffer (1 liter):

500 mM  $\text{NaH}_2\text{PO}_4$                       69.0 g  $\text{NaH}_2\text{PO}_4 \cdot \text{H}_2\text{O}$  (MW 137.99 g/mol)

1.5 M NaCl                                      87.7 g NaCl (MW 58.44 g/mol)

Adjust pH to 8.0 using NaOH.

### Buffers for purification of proteins using *Strep-Tactin Superflow*

#### Buffer NP\* (1 liter):

50 mM  $\text{NaH}_2\text{PO}_4$                       6.90 g  $\text{NaH}_2\text{PO}_4 \cdot \text{H}_2\text{O}$  (MW 137.99 g/mol)

300 mM NaCl                                      17.54 g NaCl (MW 58.44 g/mol)

Adjust pH to 8.0 using NaOH.

#### Buffer NPD (1 liter):

50 mM  $\text{NaH}_2\text{PO}_4$                       6.90 g  $\text{NaH}_2\text{PO}_4 \cdot \text{H}_2\text{O}$  (MW 137.99 g/mol)

300 mM NaCl                                      17.54 g NaCl (MW 58.44 g/mol)

2.5 mM desthiobiotin                      0.54 g desthiobiotin (Sigma cat. no. D 1411)

Adjust pH to 8.0 using NaOH.

\* Lysis buffers for insect cells should be supplemented with 1% Igepal CA-630.

## Buffers for purification of proteins using *Strep-Tactin* Magnetic Beads

### Buffer NP-T (1 liter):

50 mM NaH <sub>2</sub> PO <sub>4</sub>	6.90 g NaH <sub>2</sub> PO <sub>4</sub> ·H <sub>2</sub> O (MW 137.99 g/mol)
300 mM NaCl	17.54 g NaCl (MW 58.44 g/mol)
0.05% Tween 20	5 ml of a 10% Tween 20 stock solution
Adjust pH to 8.0 using NaOH.	

### Buffer NPB-T (1 liter):

50 mM NaH <sub>2</sub> PO <sub>4</sub>	6.90 g NaH <sub>2</sub> PO <sub>4</sub> ·H <sub>2</sub> O (MW 137.99 g/mol)
300 mM NaCl	17.54 g NaCl (MW 58.44 g/mol)
10 mM biotin	2.44 g biotin (Sigma cat. no. B 4501)
0.05% Tween 20	5 ml of a 10% Tween 20 stock solution
Adjust pH to 8.0 using NaOH.	

### *Strep-Tactin* Superflow Regeneration Buffer (1 liter):

50 mM NaH <sub>2</sub> PO <sub>4</sub>	6.90 g NaH <sub>2</sub> PO <sub>4</sub> ·H <sub>2</sub> O (MW 137.99 g/mol)
300 mM NaCl	17.54 g NaCl (MW 58.44 g/mol)
1 mM HABA	0.24 g HABA (Sigma cat. no. H 5126)
Adjust pH to 8.0 using NaOH.	

## Buffers for purification of proteins using Ni-NTA Superflow

### Buffer NPI-10\* (1 liter):

50 mM NaH <sub>2</sub> PO <sub>4</sub>	6.90 g NaH <sub>2</sub> PO <sub>4</sub> ·H <sub>2</sub> O (MW 137.99 g/mol)
300 mM NaCl	17.54 g NaCl (MW 58.44 g/mol)
10 mM imidazole	0.68 g imidazole (MW 68.08 g/mol)
Adjust pH to 8.0 using NaOH.	

### Buffer NPI-20 (1 liter):

50 mM NaH <sub>2</sub> PO <sub>4</sub>	6.90 g NaH <sub>2</sub> PO <sub>4</sub> ·H <sub>2</sub> O (MW 137.99 g/mol)
300 mM NaCl	17.54 g NaCl (MW 58.44 g/mol)
20 mM imidazole	1.36 g imidazole (MW 68.08 g/mol)
Adjust pH to 8.0 using NaOH.	

\* Lysis buffers for insect cells should be supplemented with 1% Igepal CA-630.

## Buffers for purification of proteins using Ni-NTA Superflow

### Buffer NPI-250 (1 liter):

50 mM NaH<sub>2</sub>PO<sub>4</sub>                      6.90 g NaH<sub>2</sub>PO<sub>4</sub>·H<sub>2</sub>O (MW 137.99 g/mol)

300 mM NaCl                            17.54 g NaCl (MW 58.44 g/mol)

250 mM imidazole                    17.00 g imidazole (MW 68.08 g/mol)

Adjust pH to 8.0 using NaOH.

### Buffer B (1 liter):

8M Urea                                    480.50 g urea (MW 60.06 g/mol)

100 mM NaH<sub>2</sub>PO<sub>4</sub>                    13.80 g NaH<sub>2</sub>PO<sub>4</sub>·H<sub>2</sub>O (MW 137.99 g/mol)

100 mM Tris-Cl                        12.10 g Tris-Cl (MW 121.10 g/mol)

Adjust pH to 8.0 using HCl.

### Buffer C (1 liter):

8M Urea                                    480.50 g urea (MW 60.06 g/mol)

100 mM NaH<sub>2</sub>PO<sub>4</sub>                    13.80 g NaH<sub>2</sub>PO<sub>4</sub>·H<sub>2</sub>O (MW 137.99 g/mol)

100 mM Tris-Cl                        12.10 g Tris-Cl (MW 121.10 g/mol)

Adjust pH to 6.3 using HCl.

### Buffer E (1 liter):

8M Urea                                    480.50 g urea (MW 60.06 g/mol)

100 mM NaH<sub>2</sub>PO<sub>4</sub>                    13.80 g NaH<sub>2</sub>PO<sub>4</sub>·H<sub>2</sub>O (MW 137.99 g/mol)

100 mM Tris-Cl                        12.10 g Tris-Cl (MW 121.10 g/mol)

Adjust pH to 4.5 using HCl.

## Buffers for purification of proteins using Ni-NTA Magnetic Agarose Beads

### PBS (1 liter):

50 mM NaH<sub>2</sub>PO<sub>4</sub>                      6.90 g NaH<sub>2</sub>PO<sub>4</sub>·H<sub>2</sub>O (MW 137.99 g/mol)

150 mM NaCl                            8.77 g NaCl (MW 58.44 g/mol)

Adjust pH to 7.2 using NaOH.

### Buffer NPI-10-T (1 liter):

50 mM NaH<sub>2</sub>PO<sub>4</sub>                      6.90 g NaH<sub>2</sub>PO<sub>4</sub>·H<sub>2</sub>O (MW 137.99 g/mol)

300 mM NaCl                            17.54 g NaCl (MW 58.44 g/mol)

10 mM imidazole                      0.68 g imidazole (MW 68.08 g/mol)

0.05% Tween 20                        5 ml of a 10% Tween 20 stock solution

Adjust pH to 8.0 using NaOH.

**Buffer NPI-20-T (1 liter):**

50 mM NaH <sub>2</sub> PO <sub>4</sub>	6.90 g NaH <sub>2</sub> PO <sub>4</sub> ·H <sub>2</sub> O (MW 137.99 g/mol)
300 mM NaCl	17.54 g NaCl (MW 58.44 g/mol)
20 mM imidazole	1.36 g imidazole (MW 68.08 g/mol)
0.05% Tween 20	5 ml of a 10% Tween 20 stock solution
Adjust pH to 8.0 using NaOH.	

**Buffer NPI-250-T (1 liter):**

50 mM NaH <sub>2</sub> PO <sub>4</sub>	6.90 g NaH <sub>2</sub> PO <sub>4</sub> ·H <sub>2</sub> O (MW 137.99 g/mol)
300 mM NaCl	17.54 g NaCl (MW 58.44 g/mol)
250 mM imidazole	17.00 g imidazole (MW 68.08 g/mol)
0.05% Tween 20	5 ml of a 10% Tween 20 stock solution
Adjust pH to 8.0 using NaOH.	

**Buffer B-T (1 liter):**

8M Urea	480.50 g urea (MW 60.06 g/mol)
100 mM NaH <sub>2</sub> PO <sub>4</sub>	13.80 g NaH <sub>2</sub> PO <sub>4</sub> ·H <sub>2</sub> O (MW 137.99 g/mol)
100 mM Tris-Cl	12.10 g Tris-Cl (MW 121.10 g/mol)
0.05% (v/v) Tween 20	5 ml of a 10% (v/v) Tween 20 stock solution
Adjust pH to 8.0 using HCl.	

**Buffer C-T (1 liter):**

8M Urea	480.50 g urea (MW 60.06 g/mol)
100 mM NaH <sub>2</sub> PO <sub>4</sub>	13.80 g NaH <sub>2</sub> PO <sub>4</sub> ·H <sub>2</sub> O (MW 137.99 g/mol)
100 mM Tris-Cl	12.10 g Tris-Cl (MW 121.10 g/mol)
0.05% (v/v) Tween 20	5 ml of a 10% (v/v) Tween 20 stock solution

Adjust pH to 6.3 using HCl.

**Buffer E-T (1 liter):**

8M Urea	480.50 g urea (MW 60.06 g/mol)
100 mM NaH <sub>2</sub> PO <sub>4</sub>	13.80 g NaH <sub>2</sub> PO <sub>4</sub> ·H <sub>2</sub> O (MW 137.99 g/mol)
100 mM Tris-Cl	12.10 g Tris-Cl (MW 121.10 g/mol)
0.05% (v/v) Tween 20	5 ml of a 10% (v/v) Tween 20 stock solution
Adjust pH to 4.5 using HCl.	

## Appendix C: Regeneration of *Strep-Tactin Superflow*

*Strep-Tactin Superflow* can be regenerated according to the following procedure. The resin can be regenerated a maximum of two times and should then be discarded.

- 1. Wash the column three times with 5 column volumes *Strep-Tactin* Regeneration Buffer.**  
*Strep-Tactin* Regeneration Buffer contains HABA (4-hydroxyazobenzene-2-carboxylic acid). The color change from white to red indicates that the column has been regenerated by displacement of desthiobiotin.
- 2. Wash the column twice with 4 column volumes of *Strep-Tactin Superflow* Lysis Buffer.**
- 3. Store *Strep-Tactin Superflow* resin in *Strep-Tactin Superflow* Lysis Buffer at 4°C.**

## Ordering Information

Product	Contents	Cat. No.
<i>Strep</i> -Tactin Superflow Cartridge (1 ml)	For FPLC purification of <i>Strep</i> -tagged proteins	30120
<i>Strep</i> -Tactin Superflow (2 ml)*	For batch and FPLC purification of <i>Strep</i> -tagged proteins: 2 ml <i>Strep</i> -Tactin-charged resin (max. pressure: 140 psi)	30001
<i>Strep</i> -Tactin Magnetic Beads (2 x 1 ml)*	For micro-scale purification of <i>Strep</i> -tagged proteins: 2 x 1 ml <i>Strep</i> -Tactin-charged magnetic agarose beads (10% suspension)	36311
<i>Strep</i> -tag Antibody (100 µg)	Mouse monoclonal antibody that recognizes the <i>Strep</i> -tag II epitope; lyophilized, for 1000 ml working solution	34850
pQE-TriSystem <i>Strep</i> Vector	pQE-TriSystem <i>Strep</i> vector, 25 µg	33913
His- <i>Strep</i> pQE-TriSystem Vector Set	pQE-TriSystem His- <i>Strep</i> 1 and pQE-TriSystem His- <i>Strep</i> 2 vectors, 25 µg each	32942
<b>Related products</b>		
EasyXpress Linear Template Kit Plus (20)	For generation of expression constructs using 2-step PCR, 20 reactions	32723
EasyXpress Protein Synthesis Mini Kit*	For 20 x 50 µl in vitro synthesis reactions using <i>E. coli</i> lysates	32502
Ni-NTA Superflow Cartridges (5 x 1 ml)*	5 Cartridges pre-filled with 1 ml Ni-NTA Superflow: For FPLC purification of His-tagged proteins	30721
Ni-NTA Superflow (25 ml)*	For batch and FPLC purification of 6xHis-tagged proteins: 25 ml nickel-charged resin (max. pressure: 140 psi)	30410
Ni-NTA Magnetic Agarose Beads (2 x 1 ml)*	For micro-scale purification of 6xHis-tagged proteins: 2 x 1 ml nickel charged magnetic agarose beads (5% suspension)	36111
Penta-His Antibody, BSA-free (100 µg)*	100 µg mouse anti-(H) <sub>5</sub> (lyophilized, BSA-Free)	34660

\* Other pack sizes available; please inquire.

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