

WorkBeads 40/100 SEC

WorkBeads 40/1000 SEC

WorkBeads 40/10 000 SEC

The WorkBeads™ 40/100 SEC, WorkBeads 40/1000 SEC and WorkBeads 40/10 000 SEC resins are used for preparative size exclusion chromatography (SEC) resins in laboratory and process scale purification of proteins, virus and other biomolecules by utilizing the differences in their size. The resins are based on agarose, which is biopolymer suitable for separation of biomolecules. WorkBeads resins are cross-linked using a proprietary method that results in a very rigid structure. Although the general recommendation for SEC is to use low flow rate for best purification, the rigidity and tight particle size distribution allow purification of viruses and other large substance at high flow rate for fast processing and high yields.

- Excellent resolution and low backpressure
- Robust separation across a wide size range of proteins and other biomolecules
- Chemical stable resins that allow harsh cleaning-in-place conditions



Short protocol

This short protocol is for column packing and running WorkBeads SEC resins. Detailed instructions and recommendations for optimization are given later in this instruction. Recommended flow rates are listed in Table 1 and examples of running buffers are listed in Table 2.

1. Make a slurry of the desired resin concentration.
2. Pour the slurry into the column.
3. Pack the resin with an appropriate flow rate.
4. Apply an axial compression of less than 2%.
5. Equilibrate the column with running buffer.
6. Apply sample.
7. Elute the target protein with running buffer.
8. Wash the column with deionized water.
9. Equilibrate the column with 20% ethanol for storage.

Principle

Size Exclusion Chromatography (SEC) is a simple and reliable technique for separation of molecular components according to their size. The technique relies on a separation resin of porous beads, packed closely together in a column. The three different WorkBeads 40 SEC products have different pore size distribution, which give them different size separation ranges. The packed column is prepared by equilibration with a suitable running buffer, usually an aqueous buffer, before sample loading.

In SEC, large substances in the applied sample will elute earlier than small molecules, since large substances do not enter into the pores in the beads or they can only enter a fraction of the pores. Smaller substances can enter a larger fraction of the pores of the beads and they will therefore elute later. Intermediate size substances will enter the bead pores to different degrees, thus being eluted at different elution volumes. The eluent (buffer) that is used for equilibration of the column will enter completely into the pores of the beads. This means that the substances to be separated will be eluted in the eluent used for equilibration, thus allowing buffer exchange or salt removal (or addition). In addition to size, other characteristics e.g., shape, charge, hydrophobicity and interactions between substances or substances and resin, may affect the elution volume, thus the separation. Optimization of chromatography conditions (buffer composition, flow and sample volume) may be needed to obtain the required purification. The column is prepared for the next run by passing at least one column volume of buffer through the column. This will also ensure removal of possible remaining low molecule weight substances from the sample (e.g., salt, buffer, and low M_r impurities).

SEC is usually applied as the last purification step (polishing step) since it is limited in flow and sample volume, and since it allows removal of aggregates of the target substance, buffer exchange and salt removal.

Column packing

Follow both this general advice when packing a column and the column manufacturer's instructions. Preferably, use a column with an adjustable adaptor. In some instances, a packing reservoir or column extension may be needed. For standard SEC purification we recommend packing columns from 10x300 - 600 mm, 15x600 - 900 mm, 25x600 - 900 mm, and larger. Very large substances, e.g., viruses are eluted in the void volume and can be separated from substances (proteins and other impurities) that enter the bead pores, by using shorter columns, e.g., 200 - 300 mm bed heights.

Notice: Always make sure that the maximum pressure of the column hardware is not exceeded. Backpressure caused by the chromatography system components connected downstream of the column may reduce the maximum flow that can be used. Wear eye protection.

1. Wash the resin

The resin is provided in 20% ethanol. To avoid undue backpressure when packing, wash the desired amount of resin with several column volumes of deionized water before packing.

2. Make a slurry

Add deionized water to the washed resin to obtain a 50% to 70% slurry concentration. Make approximately 15% extra slurry to compensate for the compression during packing. The amount of slurry to be prepared can be calculated according to:

$$\text{Slurry volume} = \frac{\text{bed volume} \times 100}{\% \text{ slurry}} \times 1.15$$

3. Pour the slurry into the column

Pour the slurry slowly down the side of the column to avoid formation of air bubbles. Preferably, use a packing reservoir or an extra column tube (and connector between the columns) to extend the column volume to accommodate the entire slurry volume during packing.

4. Pack the bed

Pack the resin with a downward flow higher than the intended operational flow. We recommend 100 - 300 cm/h. A lower packing flow tend to give better column efficiency. Make sure the packing flow rate does not exceed the maximum pressure of the column hardware or the resin. The intended operational flow should not be more than 75% of the packing flow rate. For high-flow applications in shorter columns (200 - 300 mm bed height) a packing flow rate of 600 cm/h may be used.

5. Close the column

When the bed height is constant mark the bed height on the column. Stop the flow and remove the packing reservoir or extra tube. Note that the bed height will increase temporarily when the flow is stopped. If needed, adjust the bed height by removing excess resin. Be careful not to remove too much resin. Gently fill the column with packing solution to its rim without disrupting the packed bed. Open the adaptor inlet if needed to let packing solution out during insertion of the adapter on top of the packed bed. Adjust the adaptor to the mark. Apply a small axial compression of less than 2% of the final bed height by lowering the adapter further below the mark.

6. Apply a flow

Apply the operating flow of 25 - 50 cm/h or higher (e.g., 50 - 250 cm/h) in a high-flow application (taking in account section 4). Check for any gap formation above the surface of the resin bed. If a gap is observed, stop the flow and adjust the adaptor to eliminate the gap.

Purification

SEC purifications can often be done under standard recommended conditions, see recommended flow rates in Table 1 and buffer conditions in Table 2. Although this may be a good starting point for testing, optimization may be needed to obtain desired results. The purification results also depend on several other factors, such as sample complexity, resin separation range, column bed height, packing quality and sample volume.

Table 1. Recommended operating flow rates.

Resin	Recommended flow rate (cm/h)	Typical range of flow rate (cm/h)
WorkBeads 40/100 SEC	50	15 - 300
WorkBeads 40/1000 SEC	50	15 - 300
WorkBeads 40/10 000 SEC	50	15 - 300

Table 2. Recommended buffer compositions.

Buffer
1. 20 mM Na-phosphate, 150 mM NaCl, pH 7.4 (PBS)
2. 50 mM Na-phosphate buffer, pH 7.0
3. 20 mM Tris-HCl, 100 mM NaCl, pH 8.0

Equilibration

Before purification the column must be equilibrated with a suitable buffer (eluent). First the storage solution is removed by applying 0.2 column volumes (CV) of water, then with 1-2 CV of running buffer. It is recommended to monitor absorbance at 280 nm (A_{280}), conductivity and pH of the column outlet. Stable signals is a strong indication of completed equilibration. Equilibration can be done at elevated flow, although may then require a slightly bigger volume to establish equilibrium.

Sample preparation

After cell disruption or extraction, clarify the sample by centrifugation at 10 000 - 20 000 $\times g$ for 15 - 30 minutes. It is generally recommended also to pass the sample through a 0.22 - 0.45 μm filter (e.g., a syringe filter) to avoid inadvertently applying any remaining particles onto the column. If the sample contains only small amounts of particles, centrifugation may be omitted, and it is enough to only carry out filtration. Application of a sample that has not been properly clarified may reduce the performance and lifetime of the column.

Sample application

For preparative purifications it is recommended to apply sample volumes of 1 - 4 % of the column volume, e.g., up to 4 ml on a 100 ml column.

Note: We recommend *not* to use Blue Dextran as a molecular weight marker due to it may give unspecific binding to the resin.

Elution

The elution is done with the same buffer as used for equilibration. Normally, all sample components are eluted within 1 CV. If interactions between the sample components and the resin occurs, the elution volume can be larger than 1 CV. It is therefore recommended to use 1.3 CV for elution. If an additional purification will be done using the same sample and separation conditions, the column can be re-equilibrated with a small volume, e.g., 0.5 CV, before application of the next sample.

Preparation for storage

Wash the column with 0.5 CV water, then with 1.5 CV 20% ethanol or other desired storage solution.

Cleaning-in-place, CIP

After elution, apply 1 CV 1 M NaOH over 1 - 2 hours.

Optimization

The conditions used for SEC purifications may have to be optimized to give the optimal results. Below follow some recommendations.

Sample

Most sample compositions can be applied to the equilibrated column. The components of the sample will quickly move into the buffer that was used for equilibration, since the components to be separated move much faster than the buffer through the column. Very high viscosity of the sample may give zone broadening, thus reduced separation, e.g., protein concentrations of more than approx. 70 mg may negatively affect the separation. A recommendation is to slightly dilute the sample before application, but keep track of the sample volume.

Sample volume

Resolution, i.e., separation between the peaks, depends on the sample volume loaded on the column. Sample volumes of 1% - 4% of the total column volume is recommended. Smaller sample volumes 0.25% – 1 % of CV may be used to further improve resolution. Notice that application of very small amounts of sample may compromise the sample yield due to inevitable adsorption of minute amounts of material to the resin during separation. In special applications where there is a large difference in elution volume between the sample components to be separated, the sample volumes can be as much as 25% of the column volume.

Resin bed height

A longer bed height will increase the resolution. Normally a bed height of 600 - 900 mm is recommended for preparative size exclusion chromatography. For small-scale separations a 10x300 mm column may be enough for the required purification.

Flow rate

Optimization of the resolution can be carried out by lowering the flow rate. Optimum flow rate for proteins are often approx. 25 - 50 cm/h. A higher flow rate decreases the resolution. The selection of flow rate in SEC is often a trade-off between resolution and purification time. For purifications of large substances that are eluted in the void volume, a higher flow rate can often be used.

Buffer composition

It is recommended to use buffers with slightly elevated ionic strength (or conductivity, salt concentration) to mask electrostatic interactions between the resin and the substances to be separated. Addition of 150 mM NaCl to any buffer is a general recommendation. Separation can generally be done over a broad range of pH from 3 to 10. However, extreme pH and high salt concentrations may affect the structure of the proteins, or target molecules. Selection of buffer should be done within the stability window of the target substance, and to avoid any aggregation during separation.

Additives such as 1 - 10% glycerol, detergents, 0.5% arginine, or denaturants such as urea and Gua-HCl are sometimes added to the buffer for stabilisation or to keep the target soluble during separation.

Scale-up

Scale-up principles

During scale-up the ratio between sample volume and column volume should be kept constant. The column volume is scaled up by increasing the column diameter while keeping the bed height the same (e.g., 600 mm). The linear flow rate should remain the same while the volumetric flow rate increases. The volumetric flow rate for each column can be calculated according to:

$$\text{Volumetric flow rate (ml/min)} = \frac{\text{Linear flow rate (cm/h)} \times \text{Column cross sectional area (cm}^2\text{)}}{60}$$

Flow

The concepts of volumetric flow, linear flow rate and residence time is important when doing scale-up in chromatography. Volumetric flow is measured in ml/min or l/min, linear flow in cm/h and residence time in minutes. The relationship between these metrics are:

$$\text{Linear flow rate (cm/h)} = \frac{\text{Volumetric flow (ml/min)} \times 60}{\text{Column cross sectional area (cm}^2\text{)}}$$

$$\text{Residence time (minutes)} = \frac{\text{Column bed height (cm)} \times 60}{\text{Linear flow rate (cm/h)}}$$

If a smaller column has been used, the flow rate for the larger column can be calculated from the flow that was established on the small column, using the equation above by keeping the residence time of the small column the same for the larger column. This will allow an increase of the linear flow in proportion to the increase in bed height between the columns see Table 3 for examples. If the column bed heights are kept constant during scale-up the linear flow rate should be kept constant (as well as the residence time).

Table 3. Example of scale-up parameters.

Column dimension	Linear flow rate (cm/h)	Volumetric flow rate (ml/min)
10 × 300	50	0.65
25 × 300	50	4.1
25 × 600	50	4.1
25 × 900	50	4.1

Combining techniques for purification

We recommend a purification strategy based on three phases: Capture, Enhance and Polish. In the capture phase, usually one purification step, it is common to use affinity chromatography or ion exchange chromatography, to remove bulk impurities and to concentrate and stabilize the target substance. The Enhance phase (one or several purifications steps) aims at further removing impurities. The polish phase aims at removing any final impurities, and when possible adjusting the conditions of the product to be suitable for subsequent use. If the capture is performed using a selective enough method, the enhance phase may be omitted and it may be enough with a polish step.

The SEC technique is usually best applied as a polish step since it is limited in sample volume, and since it in addition to removal of impurities allows removal of co-purified aggregates of the target, and since it may allow modification of the conditions (e.g., buffer exchange or salt removal). The technique can also be useful earlier in the purification scheme in certain applications, depending on sample and purification goals.

To find out more about Bio-Works chromatography resins for additional purification, please visit www.bio-works.com

Maintenance of the resin

Unpacking and inspection

Unpack the shipment as soon as it arrives and inspect it for damage. Please, report any damage or discrepancies to your local supplier.

Cleaning

During purification, impurities such as cell debris, lipids, nucleic acids and protein precipitates from the samples may gradually build up in the resin. Fouling is typical even for well-clarified samples. The severity of this process depends on the composition of the sample applied to the column. These adsorbed impurities will reduce the performance of the packed column over time. Regular cleaning (Cleaning-in-place, CIP) keeps the resin clean, reduces the rate of further fouling, and maintains the capacity, resolution and flow properties of the packed column. Cleaning of the packed column using 1 M NaOH applied by a low flow for 2 hours or overnight is often sufficient. If possible, perform the CIP using reversed flow to release any particles derived from the sample that may have been collected on the top filter.

Sanitization (reduction of microorganisms) can be carried out using combinations of NaOH and ethanol (e.g., incubation with a mixture of 0.5 M NaOH and 40% ethanol for 3 hours). The sanitization procedure and its effectiveness will depend on the microorganisms to be removed, and needs to be evaluated for each case.

Storage

Store at 2 to 25°C in 20% ethanol.

Additional information

Product description

	WorkBeads 40/100 SEC	WorkBeads 40/1000 SEC	WorkBeads 40/10 000 SEC
Size separation range ¹	10 - 150 kD	10 - 1200 kD	10 - 10 000 kD
Exclusion limit	150 kD	1200 kD	10 000 kD
Matrix	Rigid, highly cross-linked agarose	Rigid, highly cross-linked agarose	Rigid, highly cross-linked agarose
Average particle size ² (D _{V50})	45 µm	45 µm	45 µm
Max flow rate ^{3,4}	300 cm/h (600 cm/h)	300 cm/h (600 cm/h)	300 cm/h
Chemical stability	Compatible with all standard aqueous buffers used for protein purification. Should not be stored at low pH for prolonged time.		
pH stability	2 - 13	2 - 13	2 - 13
Storage	2 to 25 °C	2 to 25 °C	2 to 25 °C

1. Globular proteins.

2. The median particle size of the cumulative volume distribution.

3. Determined in water using a 15 × 900 mm column, or 25 × 200 mm (values within brackets)

4. **Notice:** Make sure that the column hardware max pressure is not exceeded.

Intended use

WorkBeads SEC resins are intended for research and bioprocess separations.

Safety

Please read the associated Safety Data Sheets (SDS) for WorkBeads SEC resins, and the safety instructions for any equipment to be used.

Related products

Related product	Pack size ¹	Article number
Prepacked columns		
BabyBio S 1 ml	1 ml × 5	45 200 103
BabyBio Q 1 ml	1 ml × 5	45 100 103
BabyBio DEAE 1 ml	1 ml × 5	45 150 103
BabyBio Dsalt 1 ml	1 ml × 5	45 360 103
BabyBio Dsalt 5 ml	5 ml × 5	45 360 107
BabyBio Ni-NTA 1 ml	1 ml × 5	45 655 103
BabyBio Ni-NTA 5 ml	5 ml × 5	45 655 107
BabyBio A 1 ml	1 ml × 5	45 605 103
Bulk resins		
WorkBeads 40S	25 ml	40 200 001
WorkBeads 40Q	25 ml	40 100 001
WorkBeads 40 DEAE	25 ml	40 150 001
WorkBeads Protein A	10 ml	40 605 003
WorkBeads 40 Ni-NTA	25 ml	40 651 001
WorkBeads 40 Ni-IDA	25 ml	40 650 001

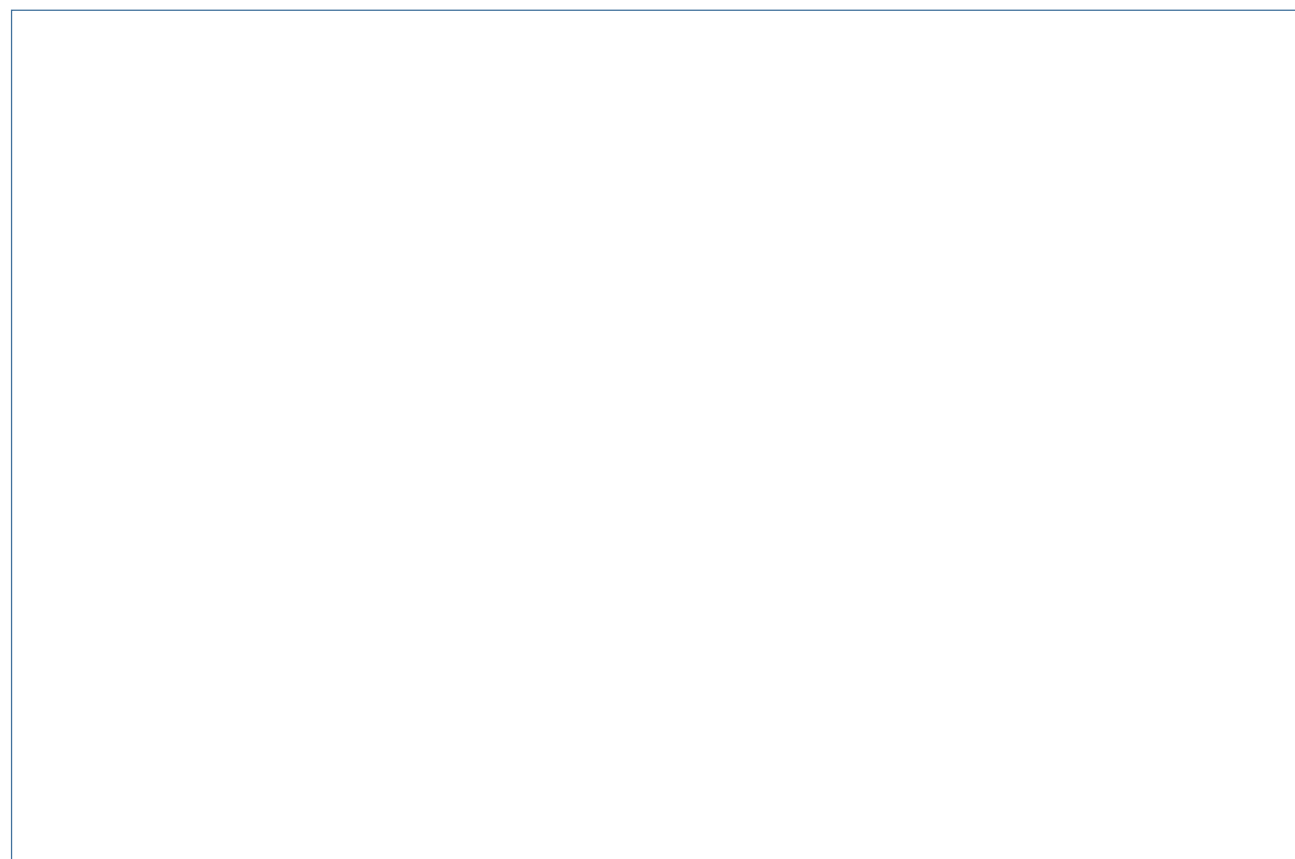
1. Other pack sizes can be found in the complete product list on www.bio-works.com

Ordering information

Product name	Pack size	Article number
WorkBeads 40/100 SEC	25 ml	40 340 001
	300 ml	40 340 003
	1 L	40 340 010
	5 L	40 340 050
WorkBeads 40/1000 SEC	25 ml	40 300 001
	300 ml	40 300 003
	1 L	40 300 010
	5 L	40 300 050
WorkBeads 40/10 000 SEC	25 ml	40 350 001
	300 ml	40 350 003
	1 L	40 350 010
	5 L	40 350 050

Orders: sales@bio-works.com or contact your local distributor.

For more information about local distributor and products please visit www.bio-works.com or contact us at info@bio-works.com



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